



# Global Methane Genetics initiative

Led by



**WAGENINGEN**  
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BEZOS  
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FUND



Global  
Methane  
Hub

## GMG Microbiome Working Group meeting

09-06-2026



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# Agenda

- Welcome
- Open discussion with all
- GMG microHub project
- Bioinformatics:
  - Boris Sepulveda (Australia)
  - Ben Perry (New Zealand)
  - Zexi Cai (Denmark)
  - Adrian López-Catalina (UK)
- Update GMG-initiative
- AOB



# Open Discussion with all

- Microbiome/Methane projects beyond GMG? Running microbiome initiatives?
- Discussion rumen vs oral vs faecal microbiome & associations with methane
- Strategies to use microbiome in breeding
- ....



Updates of GMG-microHub project

# Update GMG

## NEW projects & proposals

- **Low-methane genetics expansion to small ruminants in Africa and Asia (ICARDA)**
- **Buffalo proposal** (Lead University of Adelaide, Australia, Asian countries & South America)
- **Asia cattle proposal** (Bangladesh, India, Indonesia, and Pakistan)





**Share data,**  
*and* accelerate genetic progress  
to reduce methane emission

## Global Methane Genetics Initiative

### LOGIN

Please log in to continue to your account.

Email

Password



☐ Remember me

[Forgot password?](#)

Log in



For registration, please contact us at [methane-register@datagene.com.au](mailto:methane-register@datagene.com.au)

# GMG project:

## Framework adoption & incentivization system for genetic selection as methane mitigation tool

**Methane  
breeding value  
validation  
protocol**

Webinar  
May 11, 2026



**SWOT analysis  
trait definition  
Selection index  
Breeding objective**

Webinar  
April 13, 2026



**Review of  
policy levers  
and market  
mechanisms**

Webinar  
May 7, 2026



Session at ICAR meeting in Verona: Adoption & Incentives to Breed for Methane Mitigation  
<https://icar2026.it/meeting-programme/> & panel discussion

# Webinars & WG meetings

News  
flash\*

**Towards guidelines to validate genetic evaluations for enteric methane emissions**

Date and time  
Monday, 11 May 2026  
12:00-13:00h CET

**Latin America working group meeting**

Date and time  
Tuesday, 12 May 2026  
16:00-17:30h CET

**Beef working group meeting**

Date and time  
Monday, 18 May 2026  
13:00-14:30h CET

**Microbiome working group meeting**

Date and time  
Tuesday, 9 June 2026  
12:00-13:30h CET

**Webinar planning GMG initiative 2026**

**Measuring methane emissions using Laser Methane Detection (LMD) in rural livestock systems: key considerations**

Date and time  
Wednesday 25 March 2026  
9:00-10:30h CET

**SWOT analysis of methane trait definitions and consequences for breeding programs**

Date and time  
Monday, 13 April 2026  
12:00-13:00h CET

**Dairy working group meeting**

Date and time  
Thursday, 23 April 2026  
12:00-13:30h CET

**Sniffer data aligning and editing pipelines**

Date and time  
Wednesday, 29 April 2026  
14:00-15:30h CET

**Sheep working group meeting**

Date and time  
Thursday, 30 April 2026  
12:00-13:00h CET

**Enabling adoption of methane-reducing breeding programs through GHG accounting and MRV frameworks**

Date and time  
Thursday, 7 May 2026  
12:00-13:00h CET

**Africa working group meeting**

Date and time  
Friday, 8 May 2026  
13:00-14:30h CET

GMG newsletter:



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\* <https://www.wur.nl/en/news/methane-emissions-cows-and-sheep-can-be-reduced-25-using-breeding-programmes>

# In person meetings



Save  
the  
date



EOI Call for Papers Extended!



## Courses:

### **Methane:**

Data editing,  
Phenotypes Genetic  
analyses, Breeding  
program

### **Microbiome:**

Bioinformatics, data  
analysis, trait  
definition, breeding  
program



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# Any other business & closure

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Hub

Contact:  
[gmg@wur.nl](mailto:gmg@wur.nl)



**June 9th**

GMG presentation meeting



THE UNIVERSITY of EDINBURGH  
The Royal (Dick) School  
of Veterinary Studies



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AARHUS UNIVERSITY



## Key objectives

- The microHub project will establish a **reference population** with metagenomic and genomic data and create a **genomic evaluation** system that can be used to select the parents of the next generation with microbiome profiles that reduce enteric methane emission
- Target different **breeds** and **systems** (GMG partners & external)
- Investigate the microbiome profile as a **correlated trait** for reducing methane emissions
- Lower environmental impact through metagenomics while maintaining **rumen health**



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The Royal (Dick) School  
of Veterinary Studies

- Oscar Gonzalez Recio
- Adrian López-Catalina
- Ridwan Ahmed
- Maddi Post



- Jennie Pryce
- Boris Sepulveda
- Amanda Chamberlain
- Christy J Vander Jagt

Micro  
**HUB**



- Birgit Gredler-Grandl
- Aniek C. Bouwman



- Suzanne Rowe
- Ben Perry
- Hannah Henry
- Timothy Bilton
- John McEwan
- Fern Booker



AARHUS UNIVERSITY

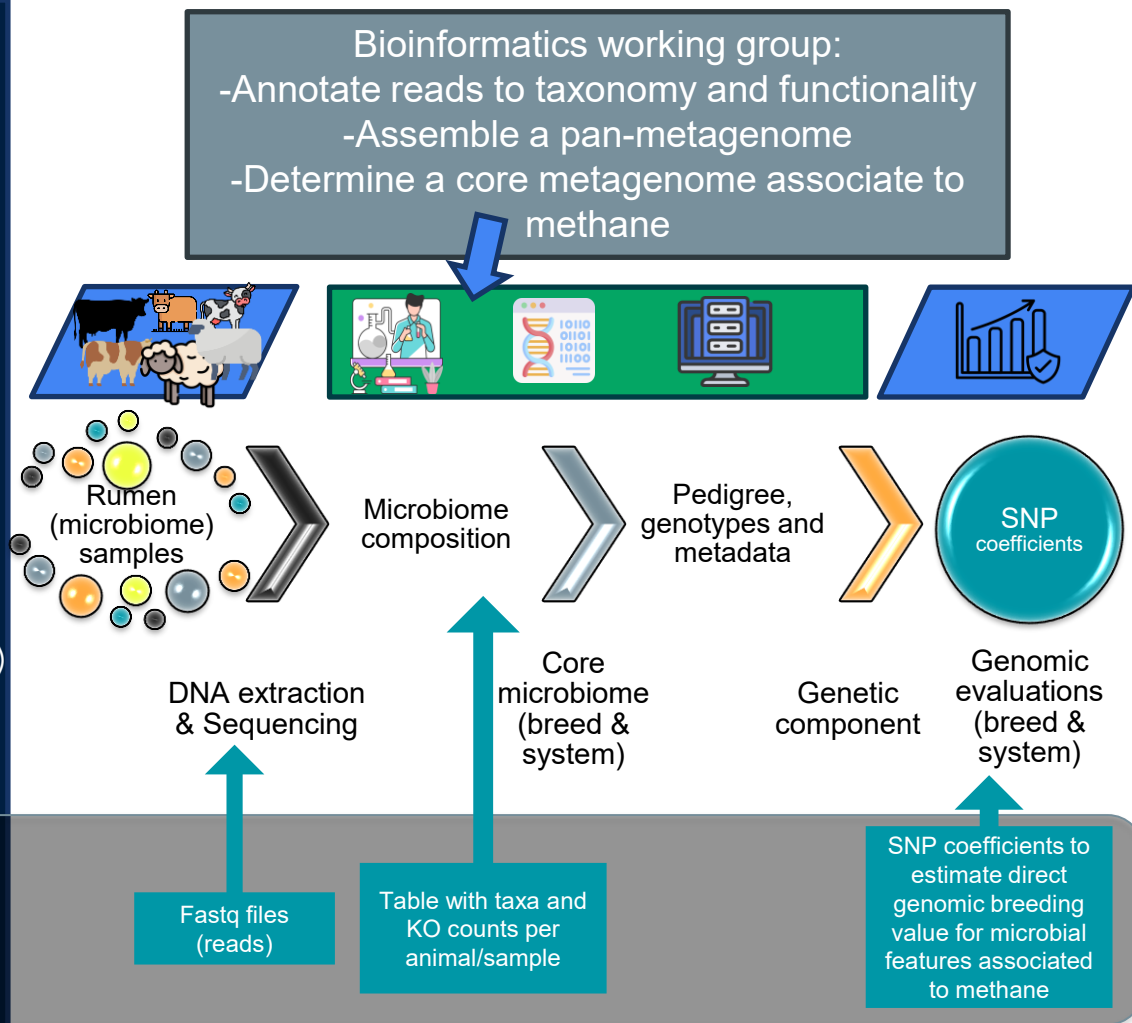
- Goutam Sahana
- Zexi Cai
- Emre Karaman
- Maria Altaf Satti

# SEQUENCING STRATEGY

- Sheep (4000)
- Dairy (7000)
- Beef (8000)
- African breeds/crosses (1200)

Information  
to be  
provided to  
partners

(from their own samples)



- Rumen metagenome as a genomic selection target to reduce enteric methane emissions

- Boris Sepulveda – Agriculture Victoria



- Restriction Enzyme-Reduced Representation Sequencing (RE-RRS) of Rumen Microbial Communities: Methane Breeding at Scale in NZ

- Ben Perry - AgResearch



- Assembly-based metagenomic profiling

- Zexi Cai – Aarhus University



- Assigning functional annotation to microbiome reads

- Adrián Lopez Catalina- The University of Edinburgh



# Open discussion and collaborations



Feel free to reach out to:

Oscar Gonzalez Recio  
[oscar.gonzalezrecio@ed.ac.uk](mailto:oscar.gonzalezrecio@ed.ac.uk)



# Rumen metagenome as a genomic selection target to reduce enteric methane emissions

**June 9th, 2026**

Boris Sepulveda  
Agriculture Victoria

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**Global  
Methane  
Genetics  
initiative**



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# TABLE OF CONTENTS



- Recent publications and Micro-HUB's aim
- Progress in Micro-HUB bioinformatics

# Recent publications

## communications biology

[Explore content](#) ▾ [About the journal](#) ▾ [Publish with us](#) ▾

[nature](#) > [communications biology](#) > [articles](#) > article

Article | [Open access](#) | Published: 13 April 2026

### Reliable enteric methane prediction from the cattle (*Bos taurus*) rumen microbiome

[Boris J. Sepulveda](#)  [Oscar González-Recio](#), [Amanda J. Chamberlain](#), [Ruidong Xiang](#), [Benjamin G. Cocks](#), [Jianghui Wang](#), [Claire P. Prowse-Wilkins](#), [Leah C. Marett](#), [S. Richard O. Williams](#), [Joe L. Jacobs](#), [Aser García-Rodríguez](#), [Jose A. Jiménez-Montero](#) & [Jennie E. Pryce](#)

[Communications Biology](#) (2026) | [Cite this article](#)



J. Dairy Sci. 108:8619–8636

<https://doi.org/10.3168/jds.2024-25436>

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### Rumen metagenome as a genomic selection target to reduce enteric methane emissions

[B. J. Sepulveda](#),<sup>1,2\*</sup>  [O. González-Recio](#),<sup>3</sup>  [A. J. Chamberlain](#),<sup>1,2</sup>  [M. Khansefid](#),<sup>1</sup>  [B. G. Cocks](#),<sup>2</sup>  [J. Wang](#),<sup>1</sup>  [C. P. Prowse-Wilkins](#),<sup>1,4</sup>  [L. C. Marett](#),<sup>5</sup>  [S. R. O. Williams](#),<sup>5</sup>  [J. L. Jacobs](#),<sup>2,5</sup>  [A. García-Rodríguez](#),<sup>6</sup>  [J. A. Jiménez-Montero](#),<sup>7</sup>  and [J. E. Pryce](#)<sup>1,2</sup> 

<sup>1</sup>Agriculture Victoria Research, AgriBio, Centre for AgriBioscience, Bundoora, VIC 3083, Australia

<sup>2</sup>School of Applied Systems Biology, La Trobe University, Bundoora, VIC 3083, Australia

<sup>3</sup>Departamento de Mejora Genética Animal, Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria (INIA-CSIC), Madrid 28040, Spain

<sup>4</sup>Department of Animal Science, University of California, Davis, CA 95616

<sup>5</sup>Agriculture Victoria Research, Ellinbank, VIC 3821, Australia

<sup>6</sup>Department of Animal Production, NEIKER-BRTA, Granja Modelo de Arkaute, 01080 Vitoria-Gasteiz, Spain

<sup>7</sup>Spanish Holstein Association (CONAFE), 28340 Madrid, Spain



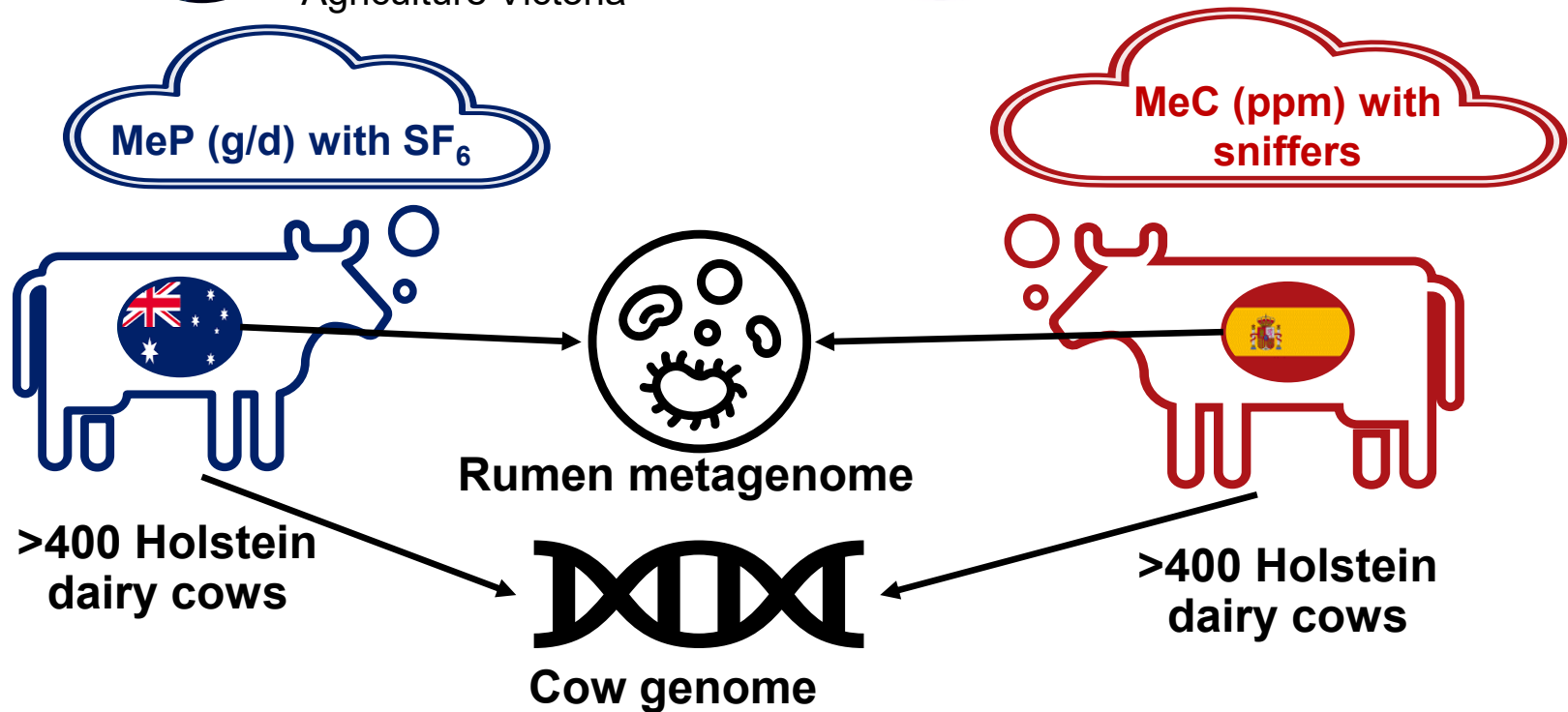
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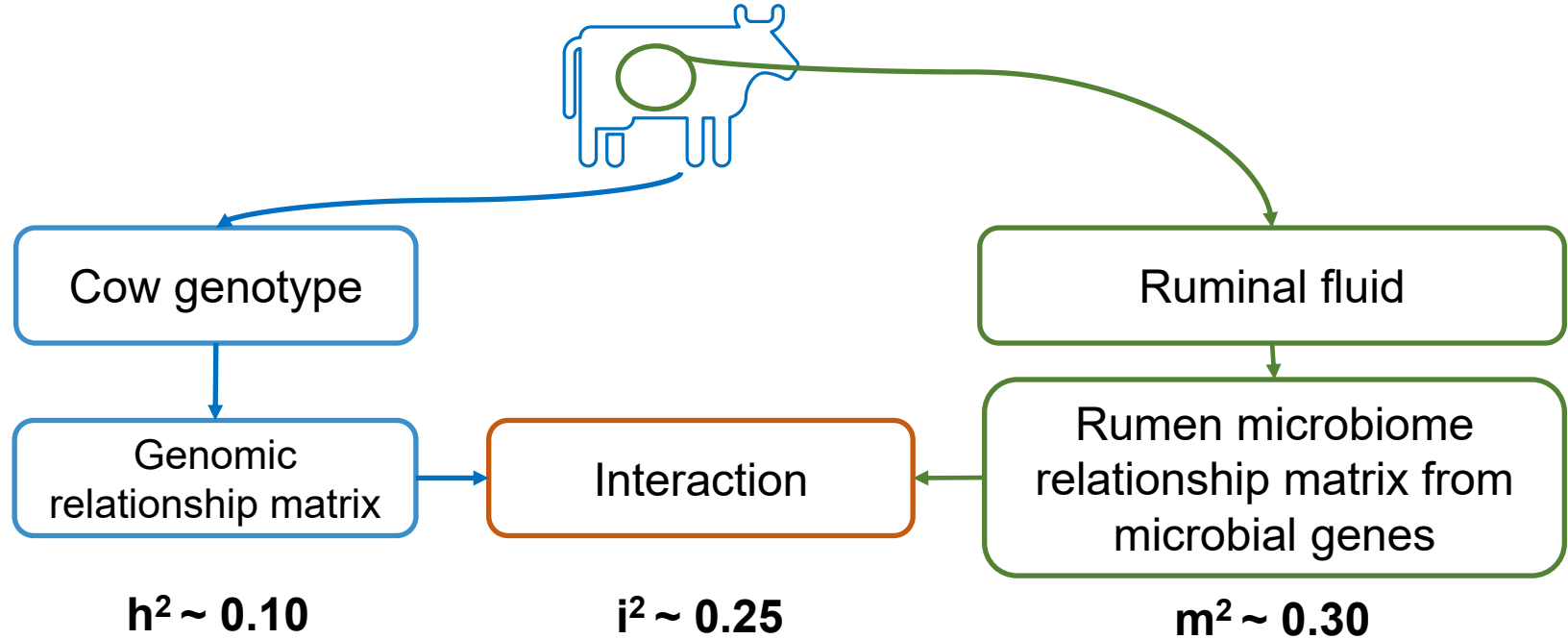
Prof Jennie Pryce  
Agriculture Victoria



Dr Oscar González Recio  
INIA - CSIC

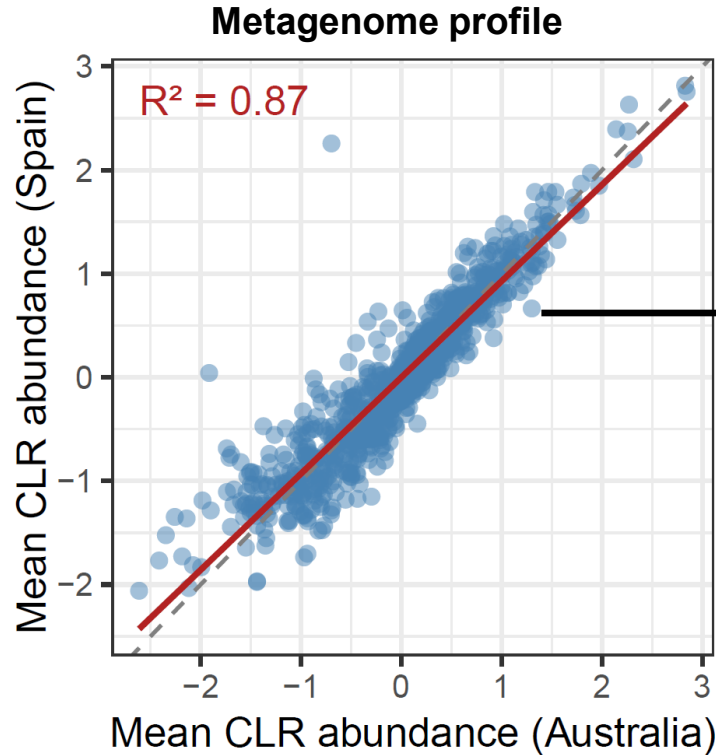


$$\text{Methane} = \mathbf{1}'\mu + \mathbf{Zg} + \mathbf{Um} + \mathbf{T(g \times m)} + \text{other effects} + \text{residual}$$

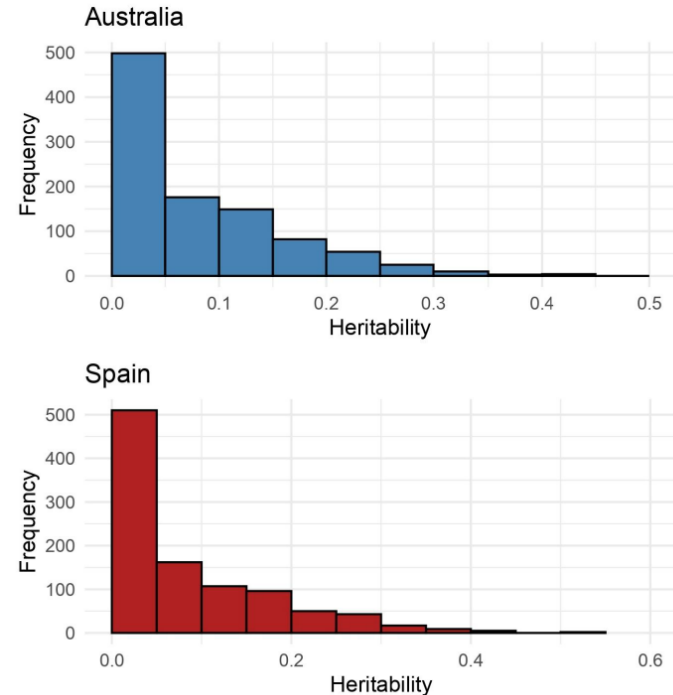


**Total variance explained: up to ~ 0.60**  
**Accuracy: up to ~0.40**

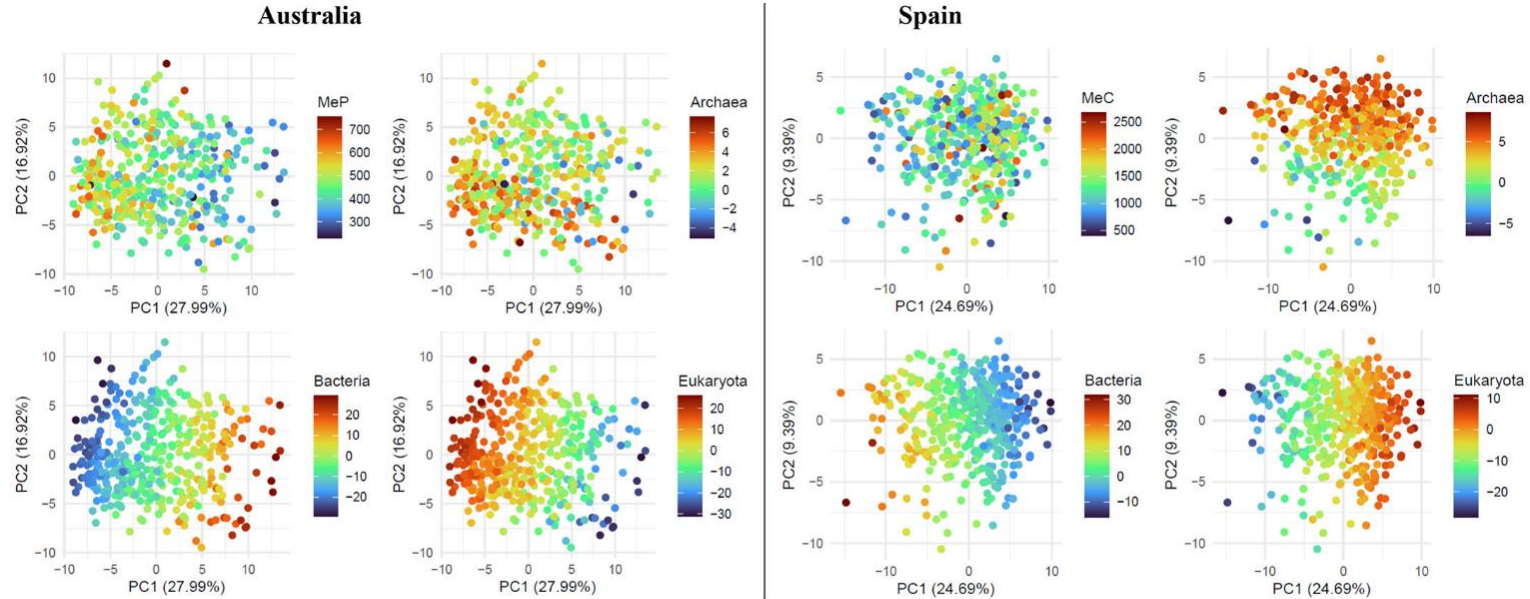
# Similarities between Australia and Spain



## Heritability ( $h^2$ ) of metagenome features



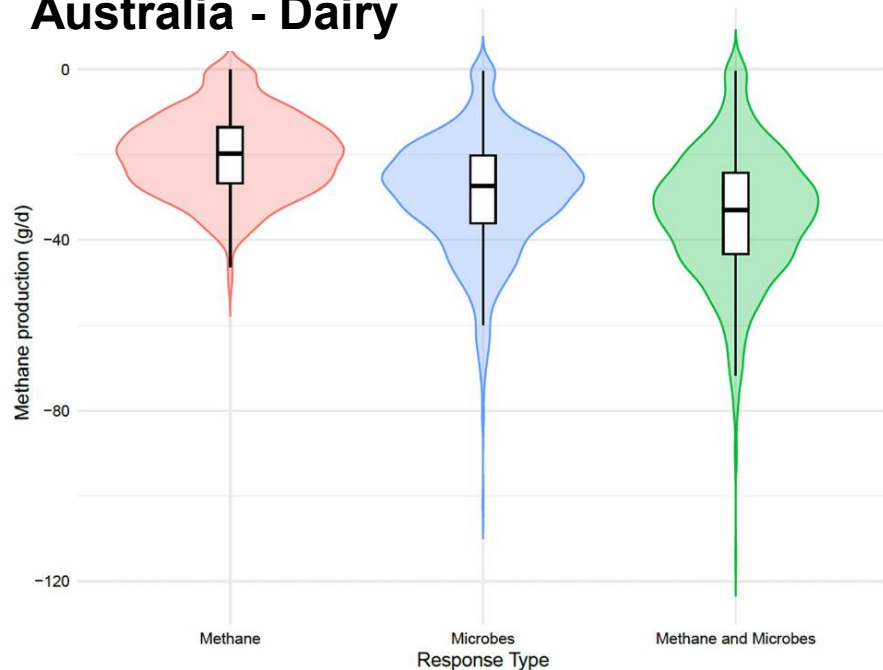
# Similarities between Australia and Spain



**Figure 1.** First 2 principal components (PC), depicting coordinates of the ruminal metagenome of dairy cattle located in Australia and Spain at genus taxonomic rank, colored by enteric methane emissions and content of *Archaea*, *Bacteria*, and *Eukaryota*. MeP = enteric methane production, recorded as grams per day (g/d) in Australia. MeC = enteric methane concentration, recorded as parts per million (ppm) in Spain.

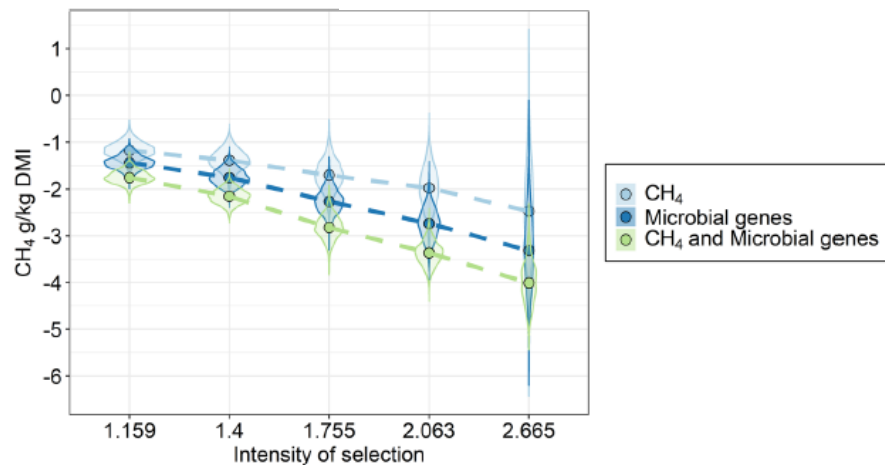
# Estimated methane reduction per generation

## Australia - Dairy



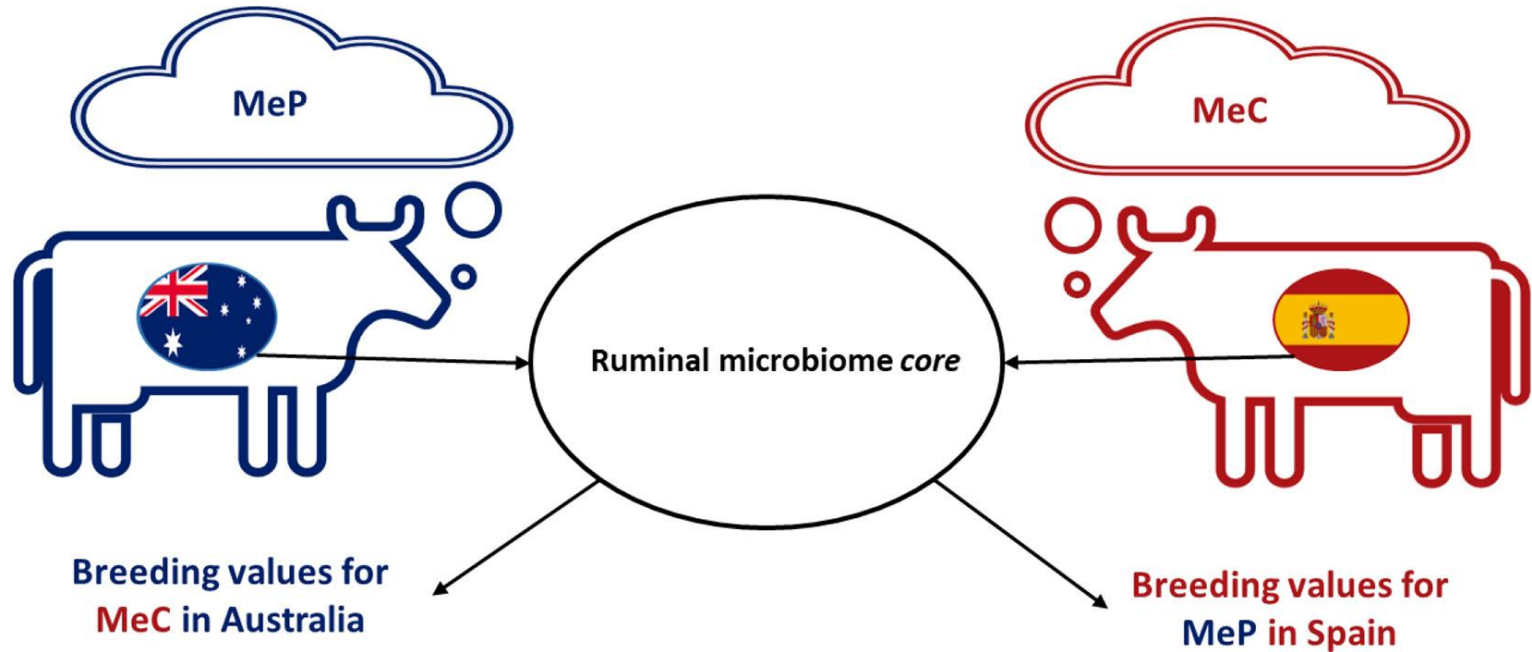
Sepulveda *et al. J. Dairy. Sci.* (2025) 08:8619–8636  
<https://doi.org/10.3168/jds.2024-25436>

## UK - Beef

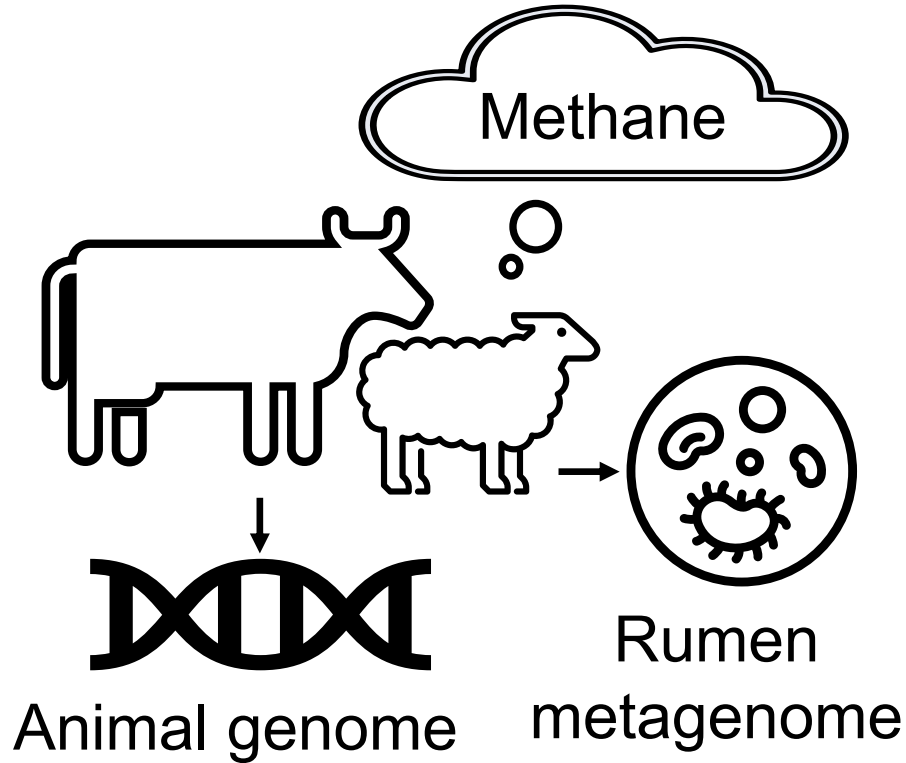


Martínez-Álvaro *et al. Commun. Bio.* 2022 Apr 12;5(1):350. doi: 10.1038/s42003-022-03293-0.

# Merging reference populations using the metagenome



# Micro-HUB

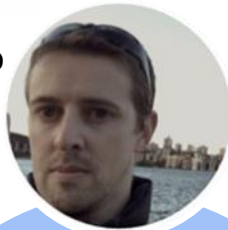


- 20K+ ruminants
- 50+ teams
- 25 countries
- Methane technologies:  
SF6, sniffers, GreenFeed,  
portable accumulation  
chambers, ...



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of Veterinary Studies

Oscar Gonzalez Recio



THE UNIVERSITY of EDINBURGH  
The Royal (Dick) School  
of Veterinary Studies

Adrian Lopez



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Aniek Bouwman



Micro-HUB  
Bioinformatics  
working group



AARHUS  
UNIVERSITY

Zexi Cai



Ben Perry



AARHUS  
UNIVERSITY

Maria Satti



Boris Sepulveda

# Bioinformatics working group

- Aims

- Unify sequencing quality control cutoffs
- Improve annotation speed
  - Annotation: Assigning biological meaning to the metagenomic DNA sequences
- Develop Standard Operating Procedures (SOPs) for bioinformatics

- Preliminary results

- Unified sequence quality control cutoffs
- Correlations  $>0.99$  between functional annotations (“genes”) obtained by different teams

- Current tasks

- Testing different approaches to improve annotation speed

# THANK YOU



## Questions?

Feel free to reach out to:

Oscar Gonzalez Recio  
[oscar.gonzalezrecio@ed.ac.uk](mailto:oscar.gonzalezrecio@ed.ac.uk)

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# **Restriction Enzyme- Reduced Representation Sequencing (RE-RRS) of Rumen Microbial Communities: Methane Breeding at Scale in NZ**

**Ben Perry, Timothy Bilton, Hannah Henry,  
Suzanne Rowe**

GMG Microbiome Working Group Meeting

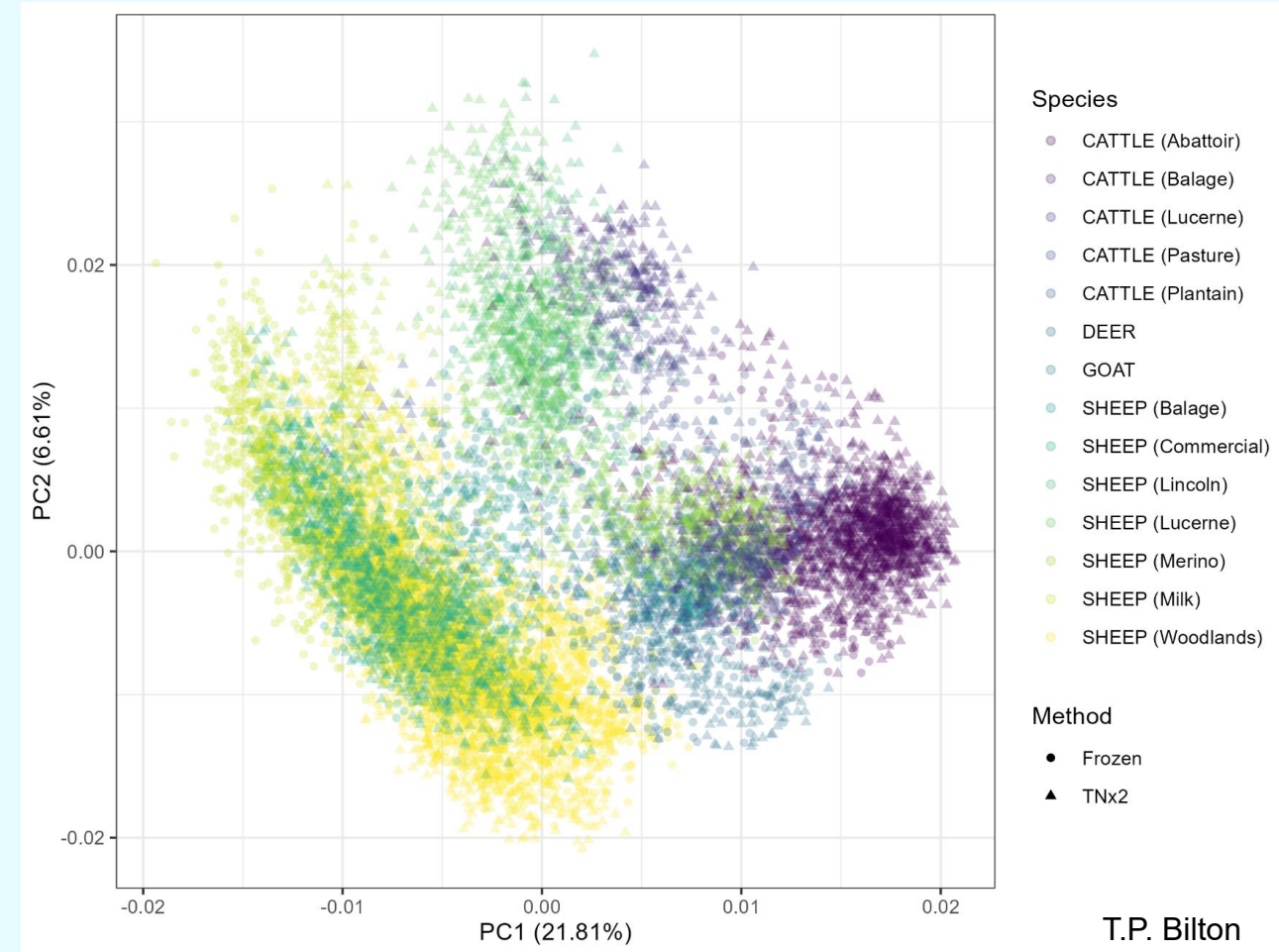
09-06-2026



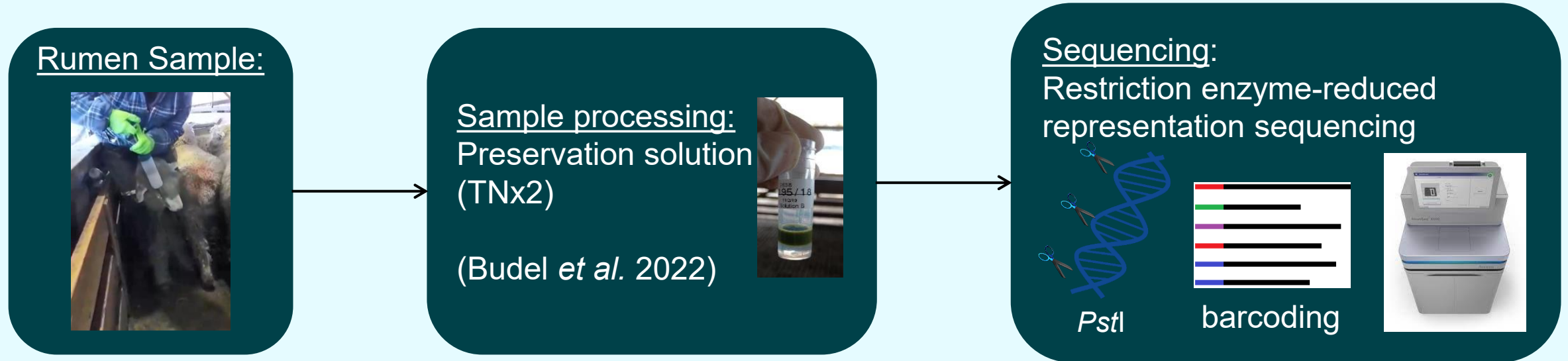
# Rumen Sampling in New Zealand – Brief Context



- We've been sampling rumen content since about 2013
- Oldest samples ~2010
- **Sheep ~ 25,200**
- Cattle ~7600
- Deer ~1400
- Develop solutions to scaling:
  - Preservant – **TNx2**
  - Sequencing – RERRSeq in Microbiomes
  - Bioinformatics – Reference and MRM based
- Emphasis on cost, throughput, CH<sub>4</sub>-prediction
- **microHUB** – Handling Sequencing and Bioinformatics Analysis of Sheep Rumen Samples



# Sample Processing and Data Generation



- Low cost RT preservative – **TNx2** mixed 1:1 with rumen content
- In-house DNA extraction and library preparation
- Illumina sequencing – NovaSeq; moving to MGI T1+ and G99 DNBSeg
- Aim for 750,00 - 500,000 reads / sample from rumen material



# High-throughput Low Cost Sequencing Restriction Enzyme-Reduced Representation Sequencing RE-RRSeq



- DNA Extraction from unfiltered rumen content
- PstI – Digestion of DNA, ligation, and size selection
  - We did try other enzymes; gives us ~1% subsampling



Red = Barcode Adapter      Green = Common adapter (New)      Brown = Paired End Sequencing Primers  
Black = DNA insert      Blue = PCR Primers (New) (i7 primers - 1 for each 768...)

5' AACTCTTTCCCTACACGACGCTCTTCCGATCTAACAATGTGCA GNNNNNNNNNNNNNNNNNNNNNNCTGCA AGATCGGAAGAGCACGTCCTGAACTCCAGTCAC 3'  
3' TGTGAGAAAGGGATGTGCTGCGAGAAGGCTAGATTGTTAC ACGTCNNNNNNNNNNNNNNNNNNNNNNGACGTTCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG 5'

AATGATACGGCGACCACCGAGATCTACACACTCTTACACTCTTTCCCTACACGACGCTCTTCCGATCT  
AACTCTTTCCCTACACGACGCTCTTCCGATCTA

TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTG (From paired end seq internet)  
TCTAGCCTTCTCGTGTGCAGACTTGAGGTCAGTGAGTGCAATAGAGCATACGGCAGAAGACGAAC

- Size selection between 220-340 bp
- 100 bp single-end reads
- In-line barcodes for large numbers (384 samples / library); ~1000s / run
- Economies of scale, in-house oligos and reagents, hit a competitive price-point

# Bioinformatic Processing: TaFeE

## Preprocessing and Reference-Based Profiling



### TaFeE — Taxonomic Feature Extraction from Microbiome Sequencing Data

A Snakemake workflow for quality control, taxonomic profiling, and functional annotation of biological sequence data (RE-RRSeq and WGS).

<https://github.com/AgResearch/TaFeE>

- We build custom Kraken2 reference indices based on the GTDB, host genomes, protists, and a small selection of fungi
- **microHUB – Unified Reference Database**

- TaFeE is our in house bioinformatics pipeline for reference-based analysis of rumen RE-RRS and shotgun metagenomic or metatranscriptomic data

### Workflow DAG

#### RE-RRSeq (RE-RRSeq-TaFeE)

```
Raw RE-RRSeq FASTQ
→ cutadapt (demultiplex)
→ BBduk (entropy/quality filter)
→ PRINSEQ++ (low-complexity filter)
→ KneadData (Trimmomatic + TRF + SILVA rRNA removal)
→ Kraken 2 (taxonomic classification)
→ taxpasta (count matrices at domain → species)
```

# Bioinformatic Processing: TaFeE Preprocessing and Reference-Based Profiling

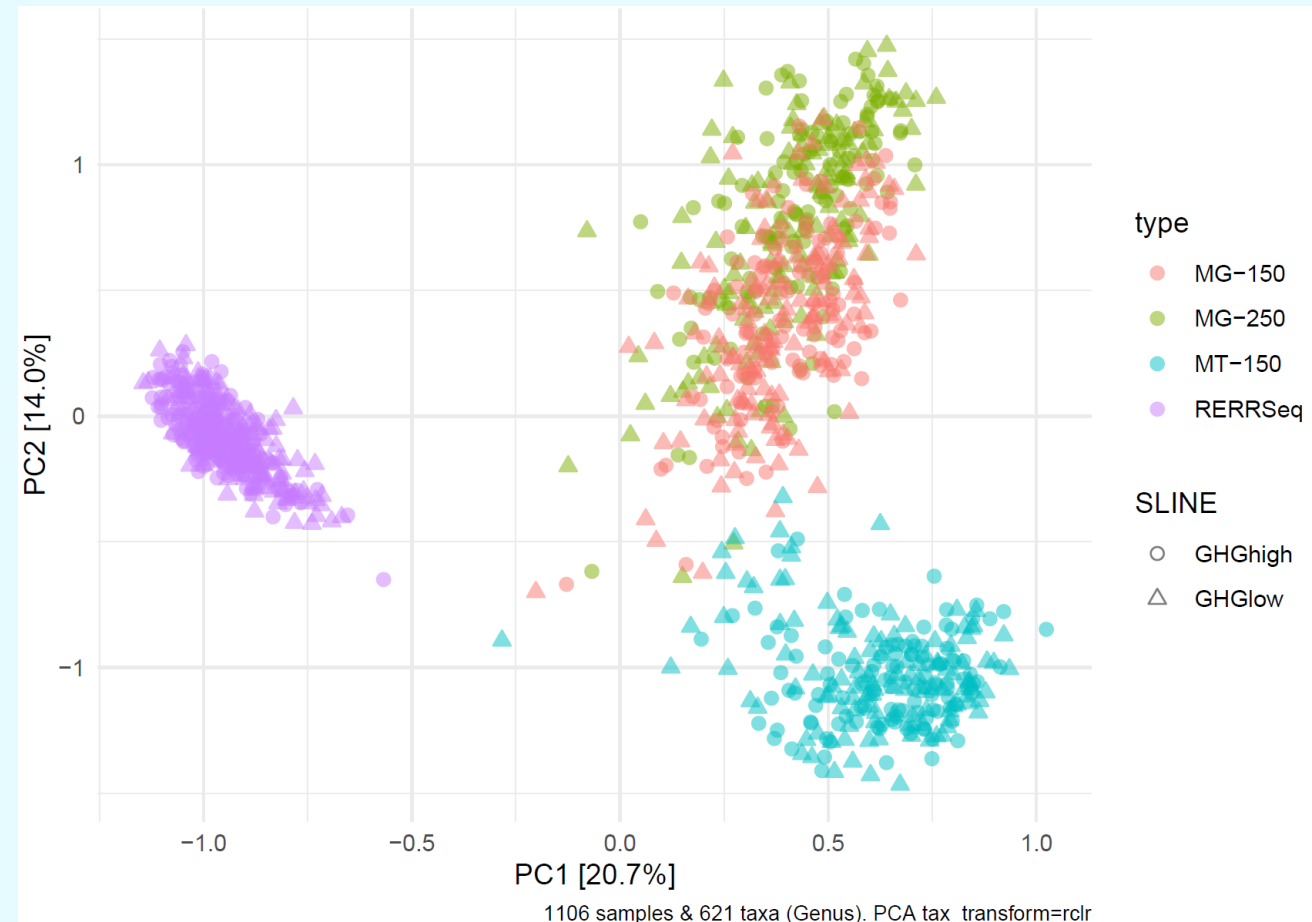


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- Unified pipeline for RE-RRSeq and shotgun metagenomics and metatranscriptomics
- Kraken2 Indices are Easily Swapped



# Bioinformatic Processing: TaFeE Preprocessing and Reference-Based Profiling

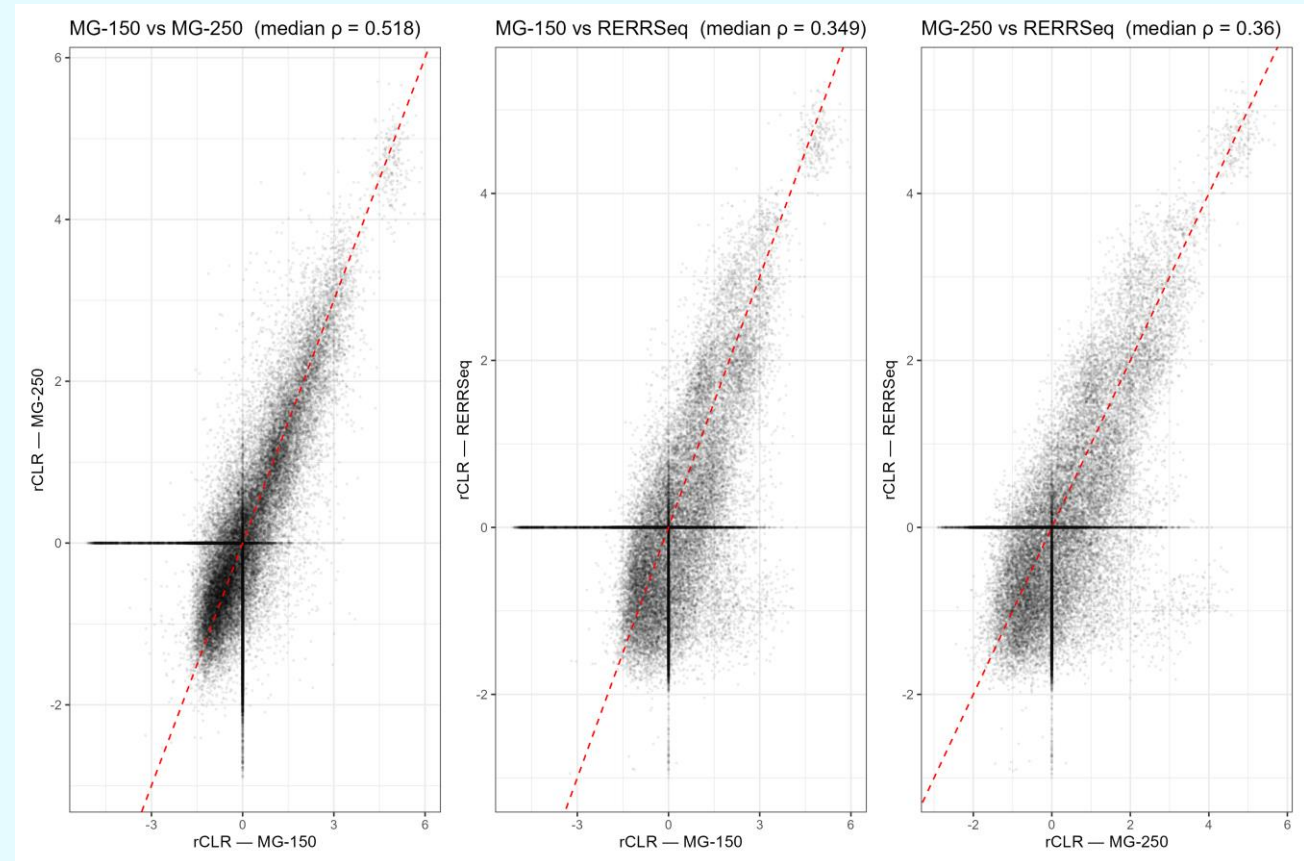


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- Unified pipeline for RE-RRSeq and shotgun metagenomics and metatranscriptomics
- Kraken2 Indices are Easily Swapped

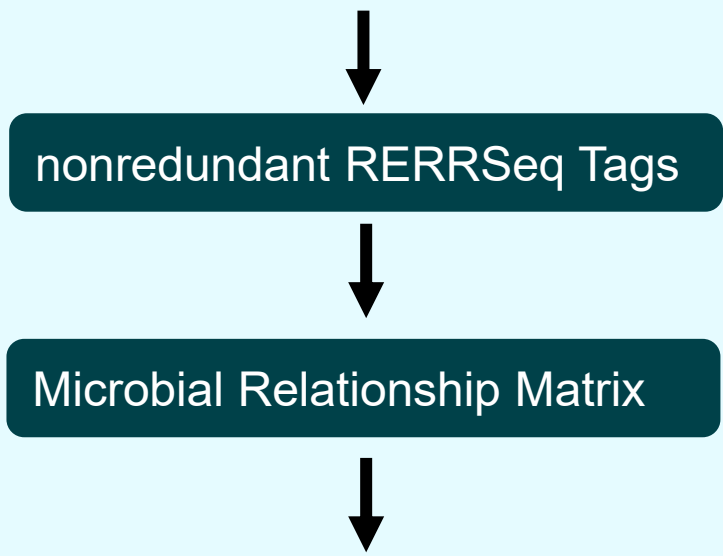


# Bioinformatic Processing: Reference-Free – MRM & Methane Modelling / Prediction

- tags are trimmed 65 bp, less than 65 bp are removed
- tags is kept if count > 0 in 25% of training set
- Resultant ~200k tags in sheep for predictions (~OTUs or ASV)
- Cohort adjustment – per tag; mean zero; variance 1 (import for predicting across farms)
- Microbial relationship matrix (Hess *et al.* 2023)

**Table 3** Heritability and microbiability estimates (standard errors)

| Trait                 | Heritability | Microbiability |
|-----------------------|--------------|----------------|
| LW8                   | 0.21 (0.023) | 0.45 (0.060)   |
| CH <sub>4</sub>       | 0.16 (0.021) | 0.54 (0.052)   |
| CH <sub>4</sub> Ratio | 0.21 (0.021) | 0.64 (0.054)   |
| CO <sub>2</sub>       | 0.21 (0.022) | 0.18 (0.046)   |



$$y = Xb + Zm + e_m$$
$$m \sim N(0, \sigma_m^2 M),$$
$$\sigma_m^2 / (\sigma_m^2 + \sigma_{e_m}^2).$$

Bilton et al. (2025) *Genet Sel Evol* **57**:25  
Bilton *et al.* (2025) *AAABG* **26**:229-232

Budel *et al.* (2022) *Anim. Microbiome* **4**:39  
Hess *et al.* (2020) *PLoS One* **15**:e0219882  
Hess *et al.* (2023) *BMC Genomics* **24**:551

# Thank you

# Presentation disclaimer



**Presentation for:**  
**GMG Microbiome Working Group Meeting**  
**2026-06-09**

## **DISCLAIMER**

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DDI: +6434899003  
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This report has been prepared by New Zealand Institute for Bioeconomy Science Limited (Bioeconomy Science Institute).

# Assembly-based metagenomic profiling

Zexi Cai

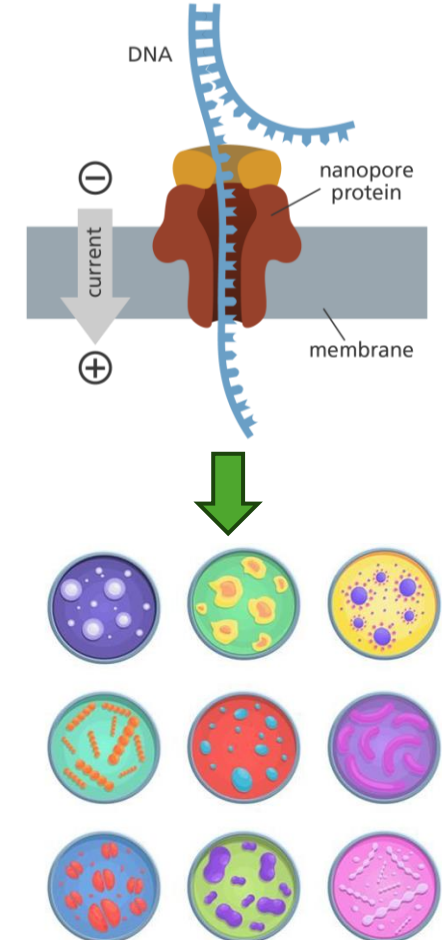
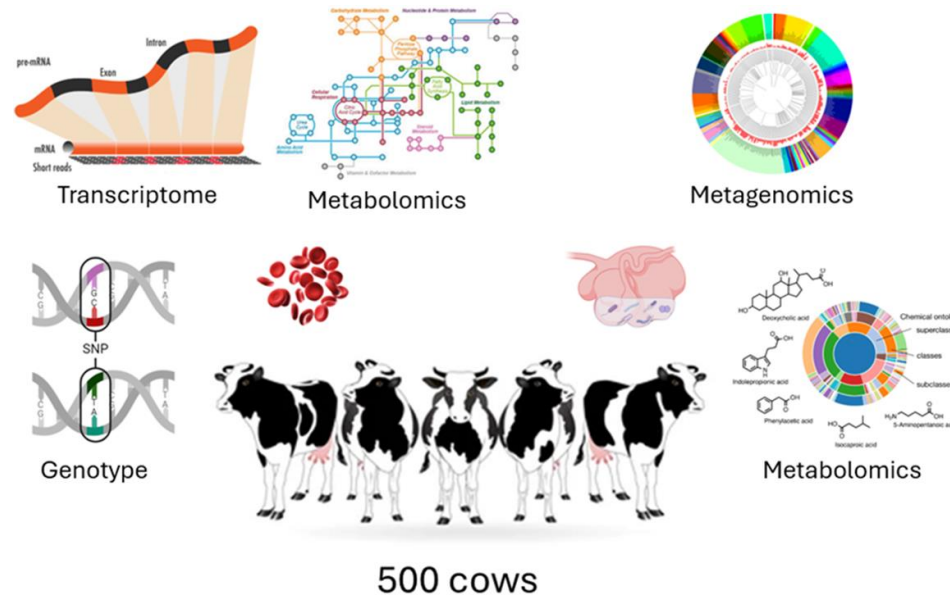
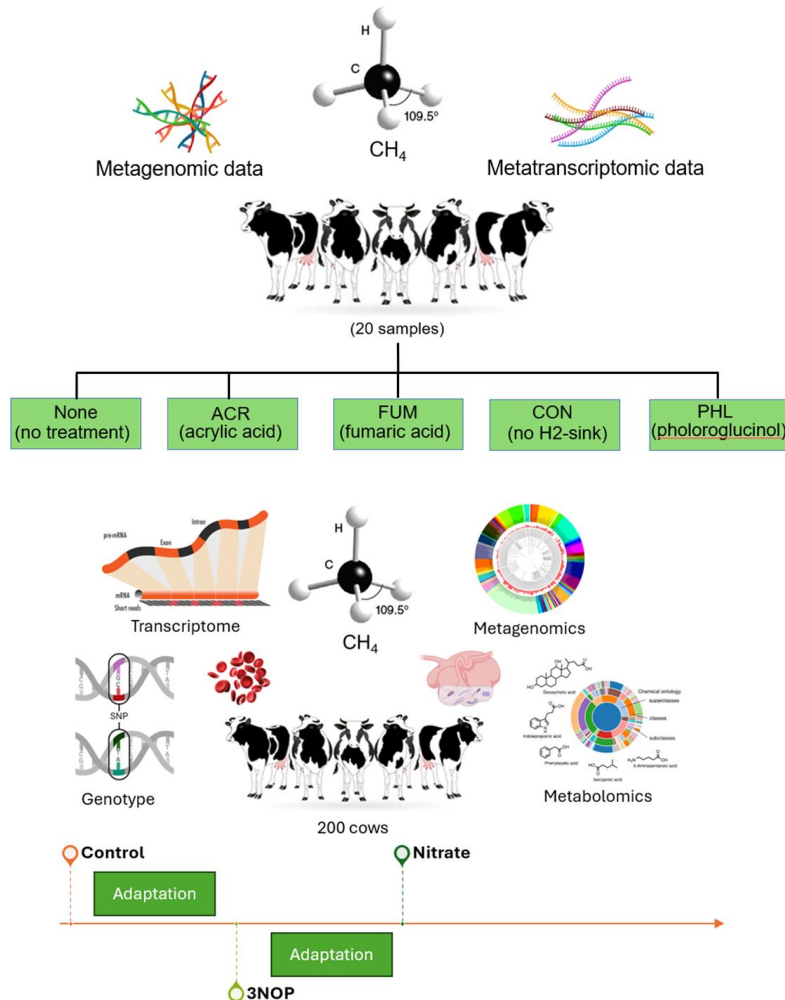
Aarhus University

# Method of choice

- Metagenomic genomes-based abundance profiling:
  - Possible to use per project information: every challenge for Marker-gene approach or protein search.
  - Possible for functional interpretation.

Metagenomic assembly -> custom database -> better taxonomy assignment

# Current activity except Microhub



MAGs is the backbone

# The Classification Gap in Metagenomics

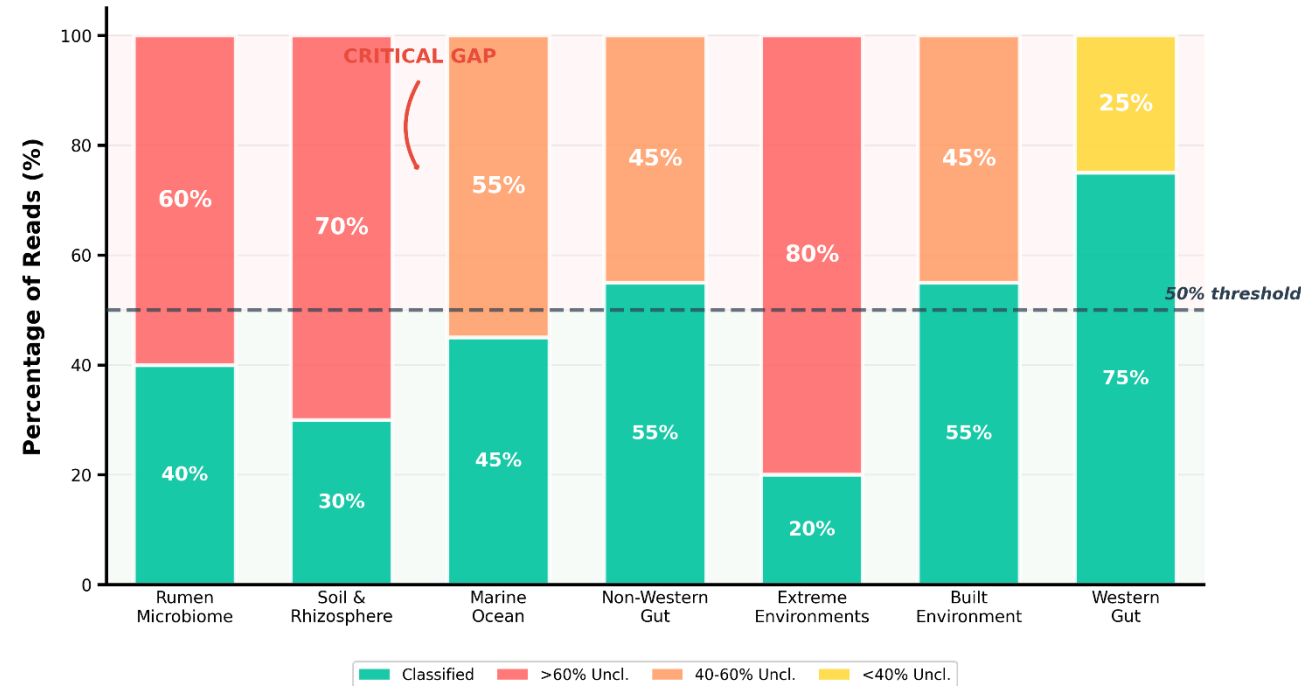
In undersampled biomes, unclassified reads exceed 50% because references skew toward cultured genomes

**Problem:** Culture bias leaves majority of microbes unidentified

**Impact:** Missed taxa, poor ecological resolution

**Solution:** Recover MAGs → verify novelty → update the reference automatically.

Classification Success Across Diverse Microbiomes



2% Microbes Cultured

98% Unknown Diversity

50% Average Unclassified

400K Genomes in GTDB

~1T Species Exist

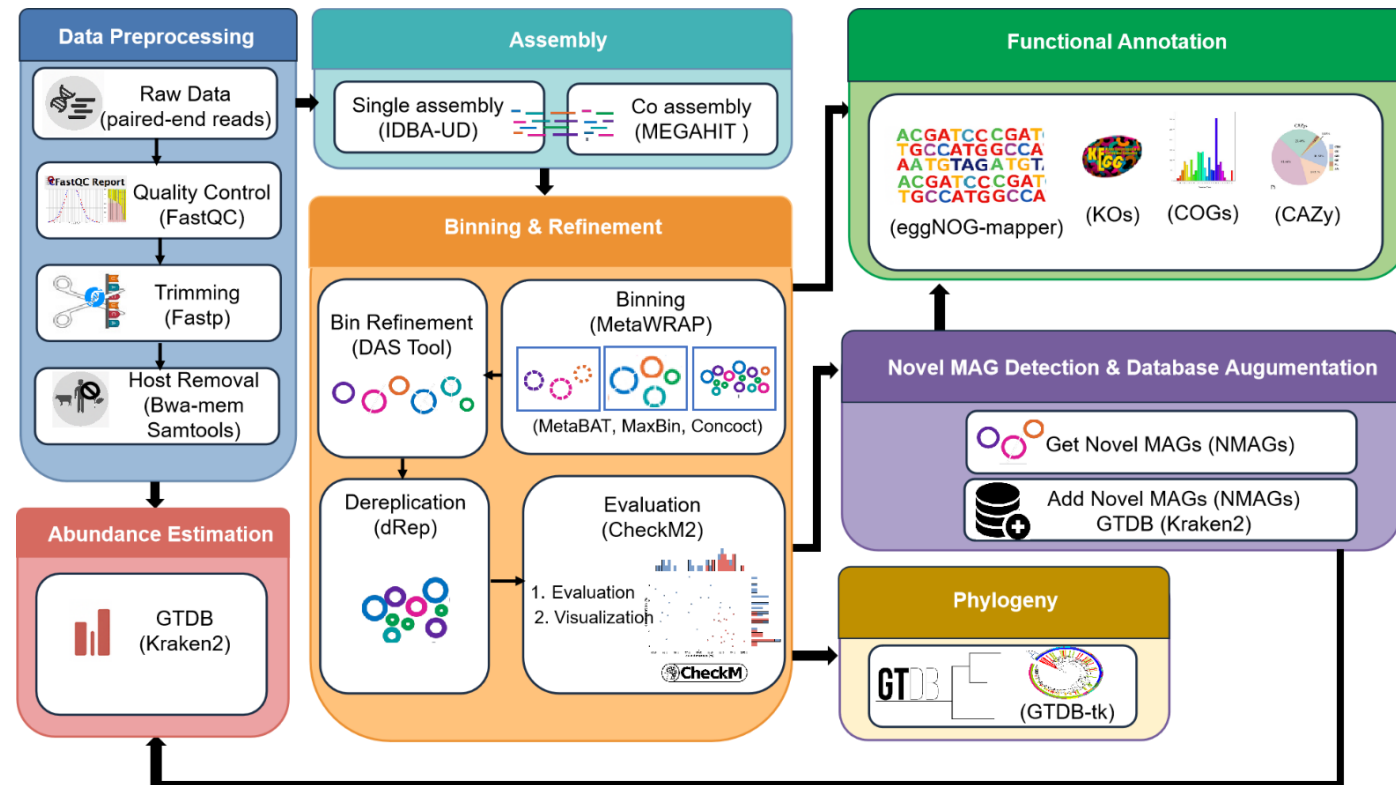
# MetaMAG Explorer



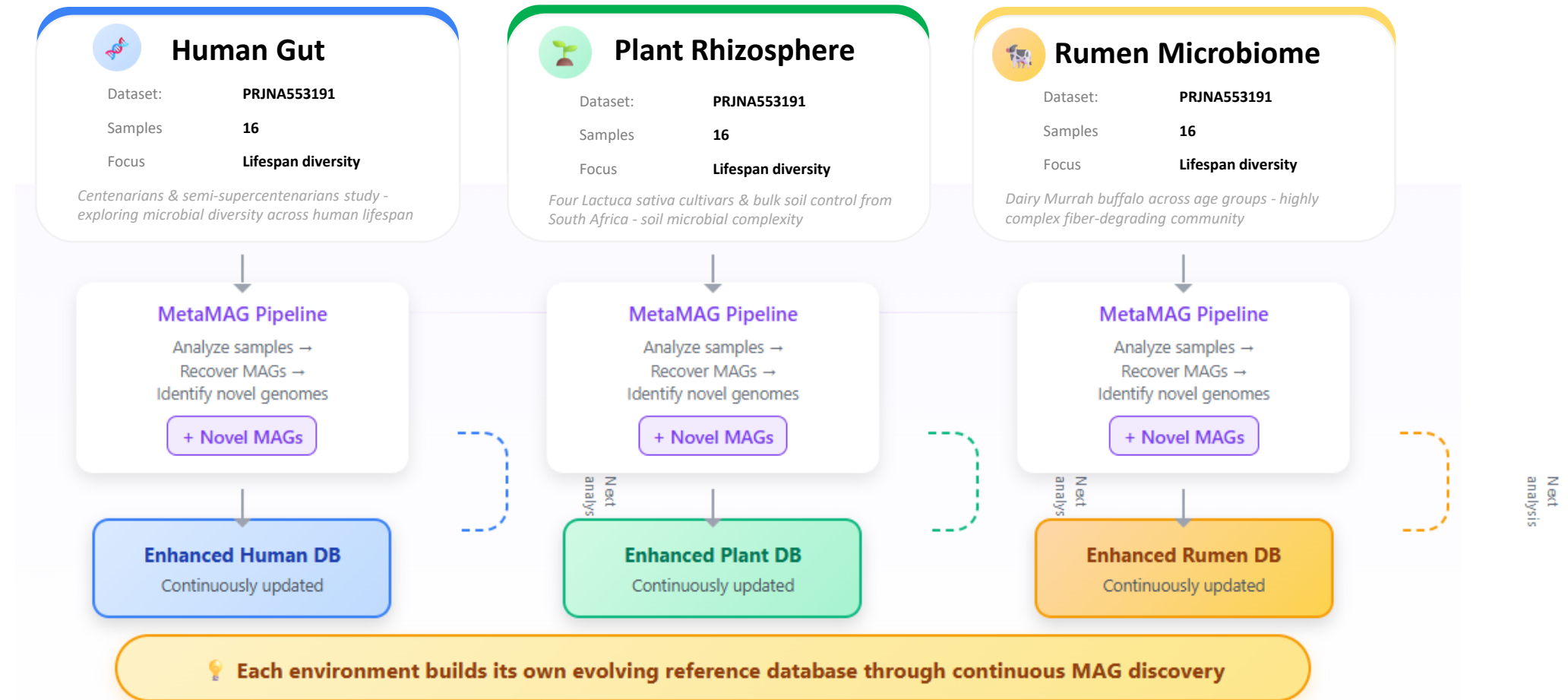
Maria Altaf Satti  
Postdoc

A modular end to end pipeline that turns raw metagenomic reads into curated MAGs and updates the reference to improve classification in under-sampled biomes.

- Automatic Database Augmentation
- Novel MAG Detection & Validation
- Rumen-Specific MAG Processing



# Pipeline Validation: Three Test Environments



# Database Augmentation Impact

**+5.4M**

Previously Unknown Reads Now Classified

**100%**

Samples Improved

**+1.27%**

Average Gain

**28**

Novel MAGs

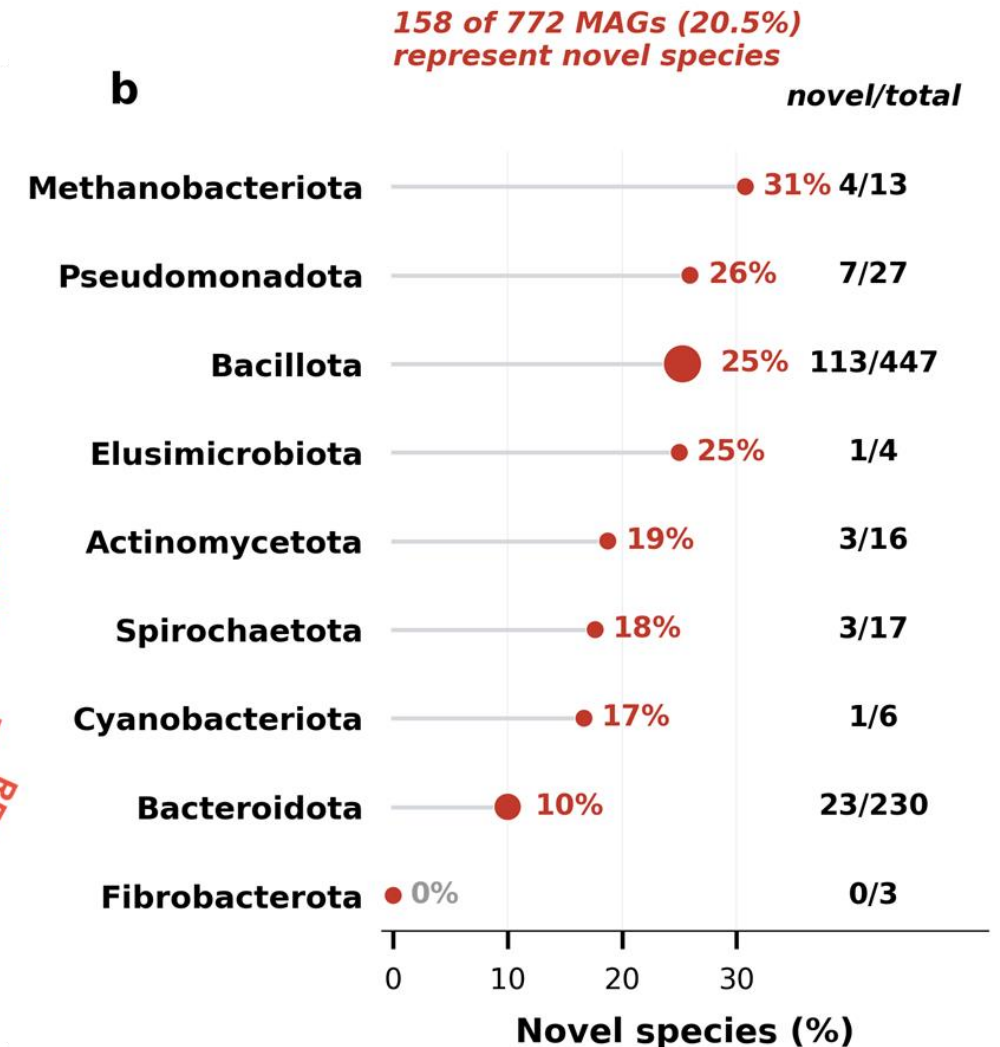
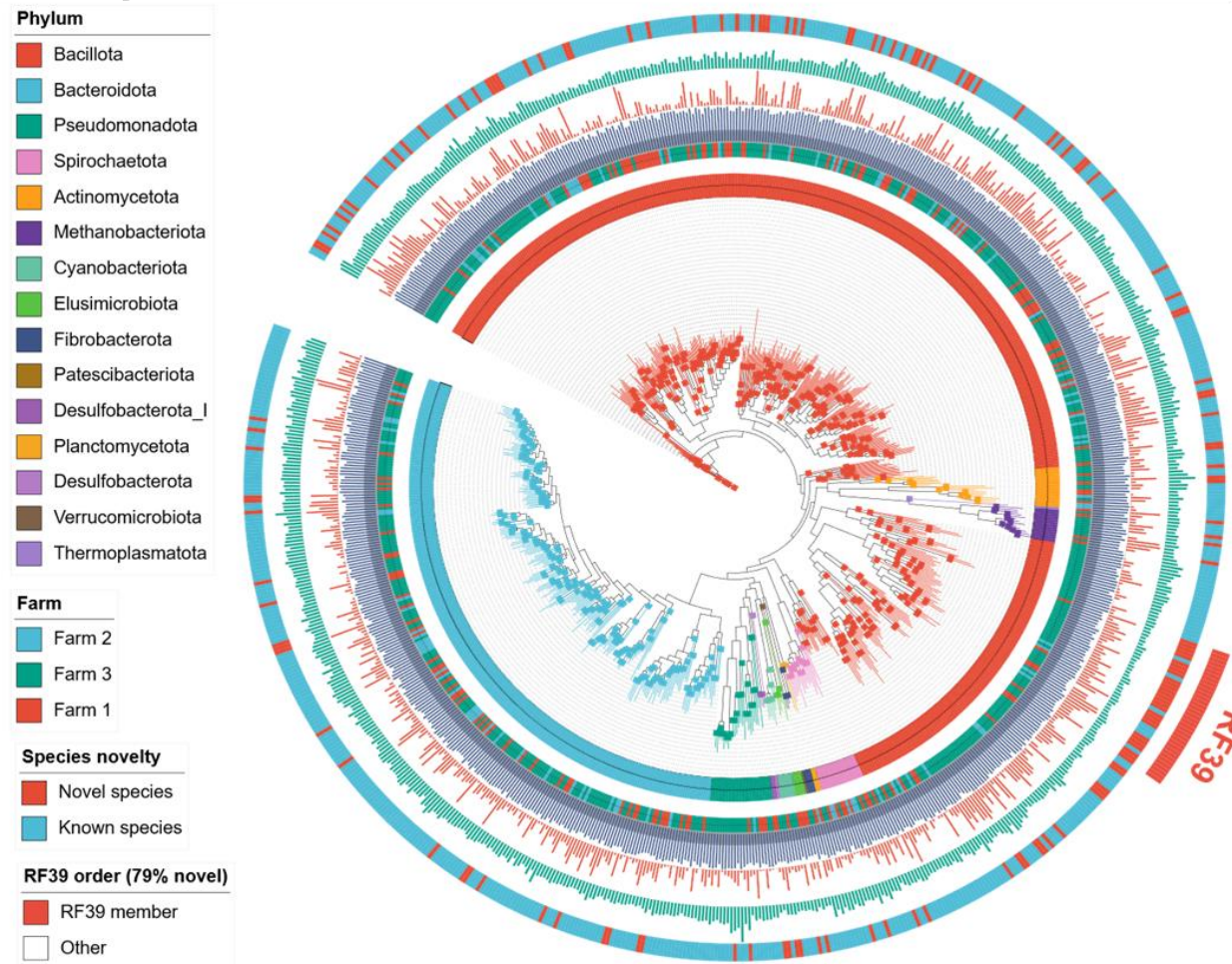
**+1.78%**

Best Result

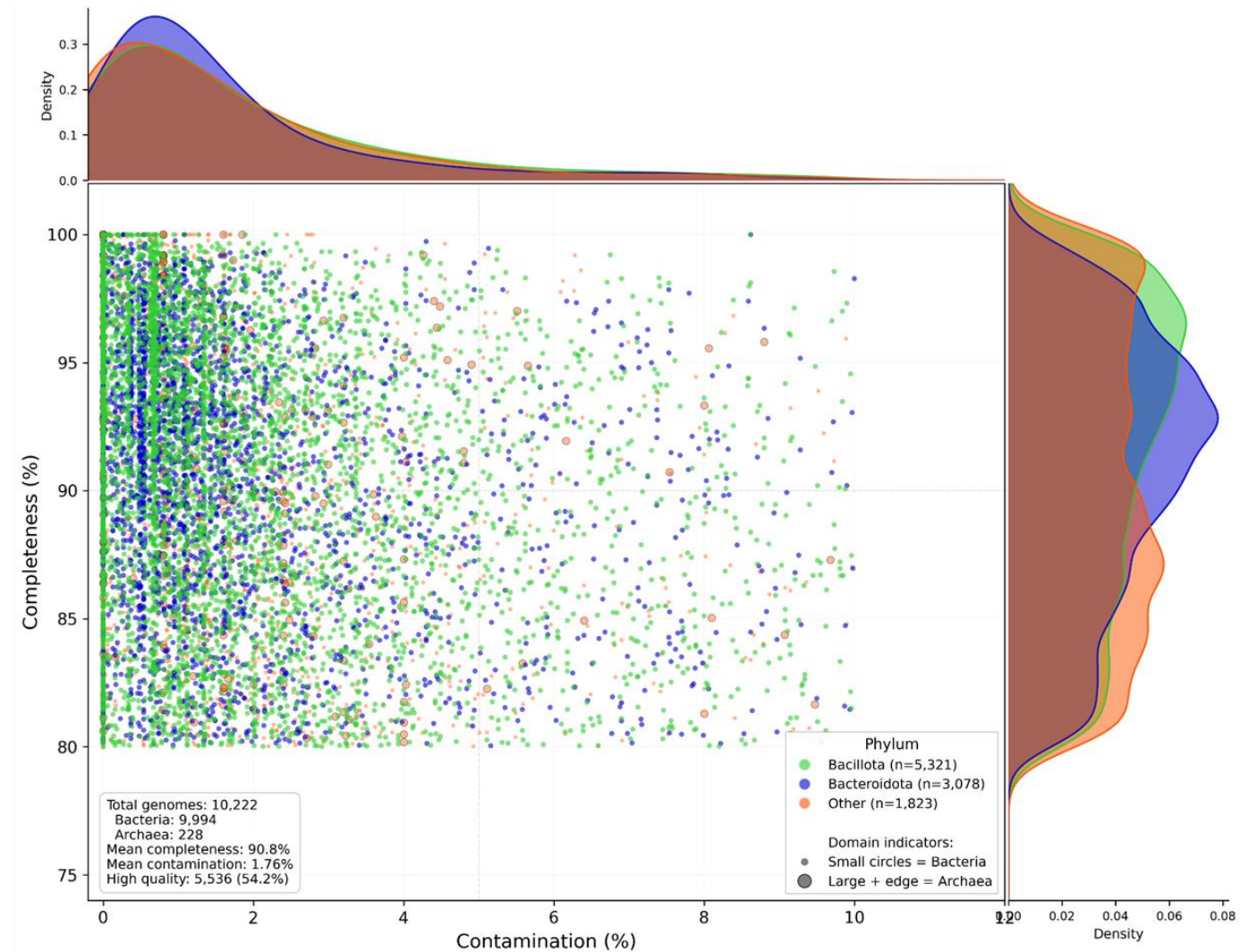
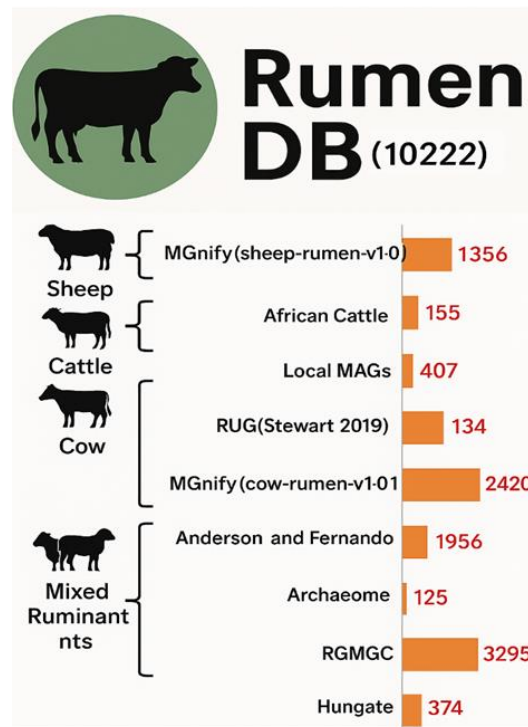
| Sample     | Original | Enhanced | Change | New Reads |
|------------|----------|----------|--------|-----------|
| Sample_962 | 74.7%    | 76.5%    | +1.78% | +522K     |
| Sample_966 | 77.0%    | 78.6%    | +1.56% | +491K     |
| Sample_955 | 78.3%    | 79.8%    | +1.54% | +419K     |
| Sample_958 | 76.9%    | 78.3%    | +1.45% | +431K     |
| Sample_964 | 76.0%    | 77.4%    | +1.39% | +410K     |

Novel MAGs found in **ALL 15 samples** – these are prevalent community members, not rare species

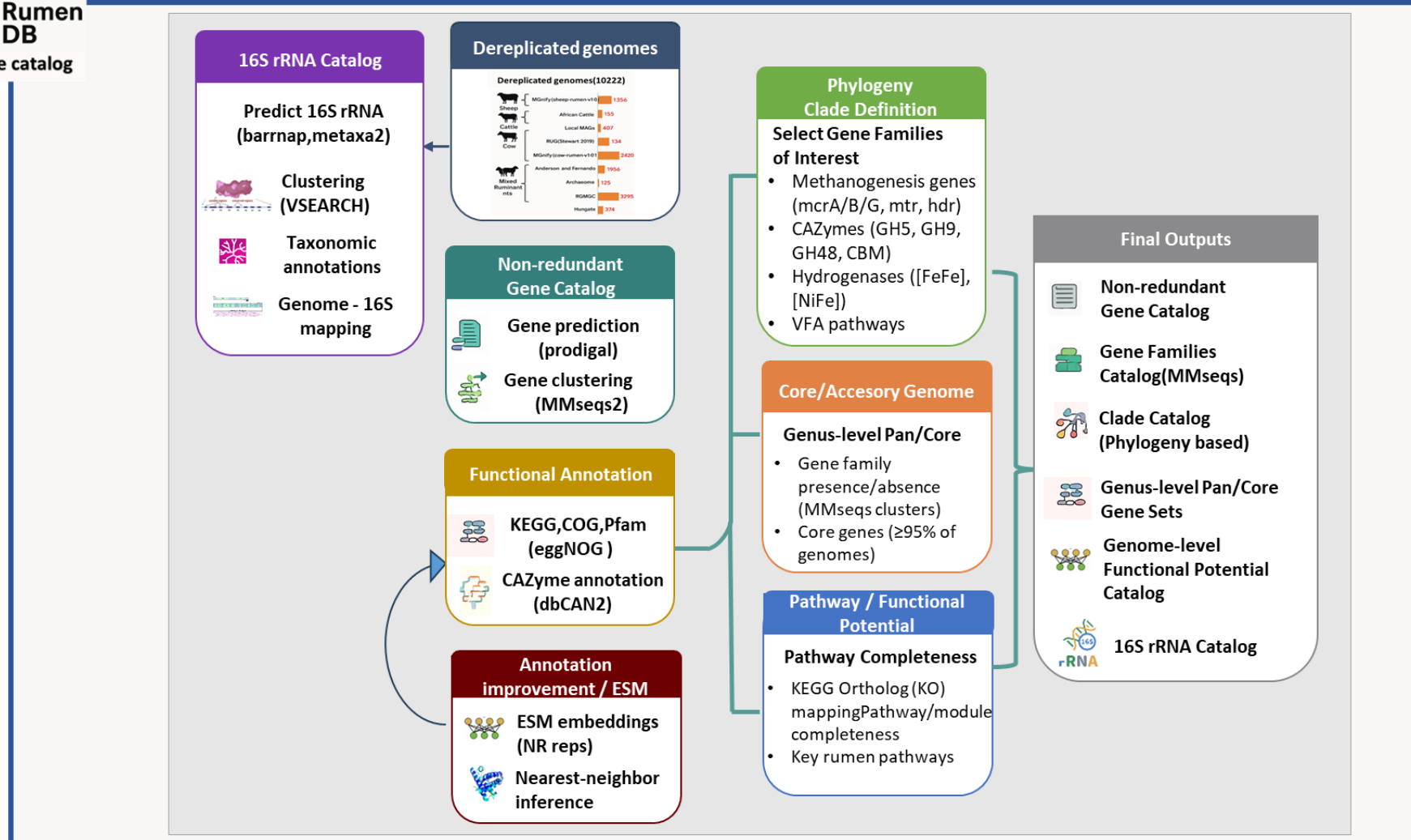
# Novel species we found across various projects



# Rumen database



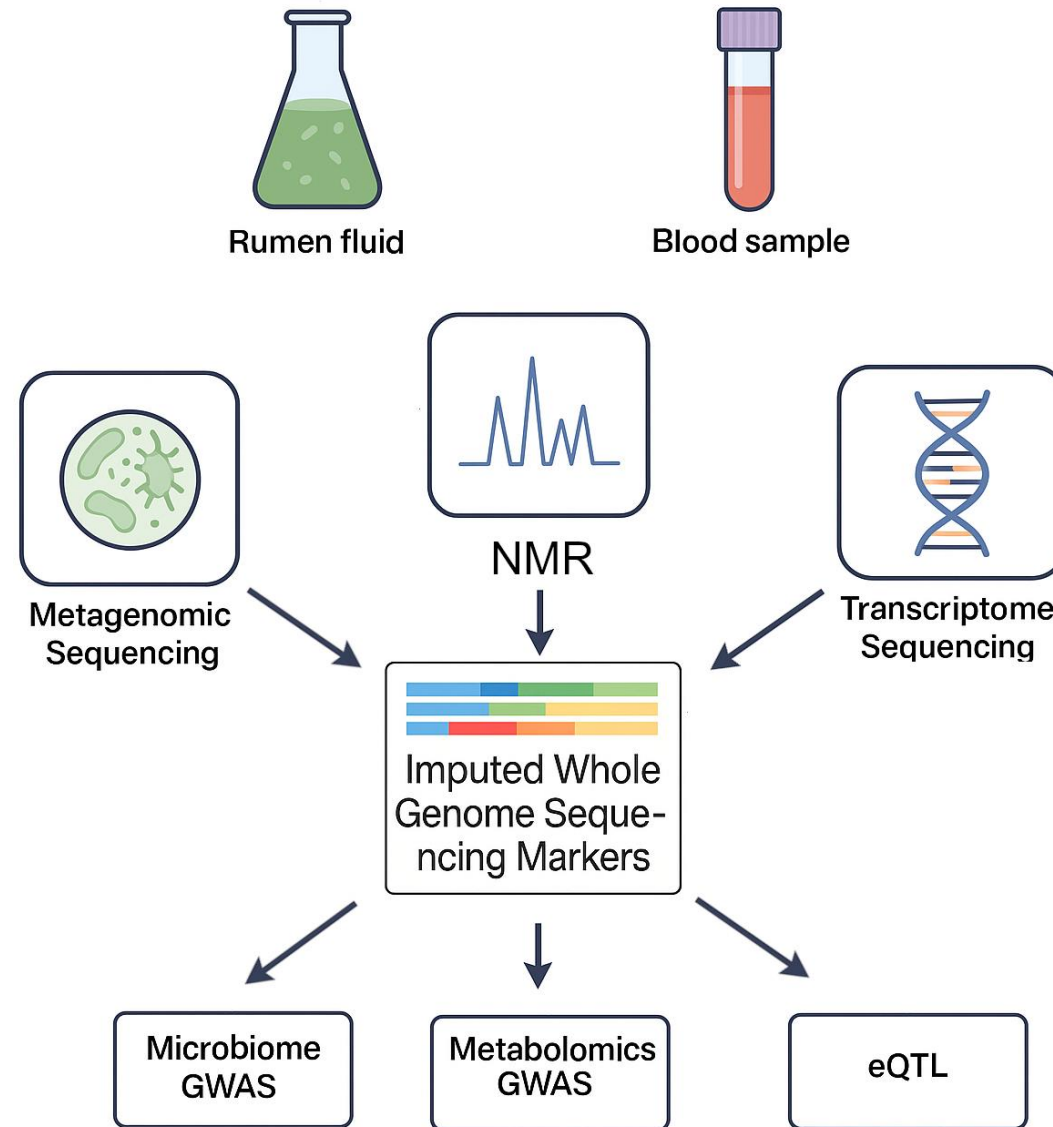
# Gene Catalog



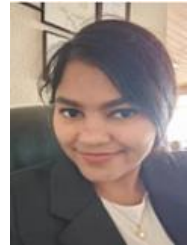
# Holo-omics to distangle the host-microbiota interaction



Sha Zhang  
Postdoc



# Deep long read sequencing for enrichment media design



Israt Farhana  
PhD student

## 4 × 4 EXPERIMENTAL DESIGN

| Cow / Period                                                                                | P1                               | P2                               | P3                               | P4                               | DATA VOLUME (Data) |                   |
|---------------------------------------------------------------------------------------------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|--------------------|-------------------|
| <br>Cow 1  | <b>3-NOP + nitrate</b><br>71 Gbp | <b>control</b><br>86 Gbp         | <b>nitrate</b><br>95 Gbp         | <b>3-NOP</b><br>97 Gbp           | Sample             | Data Volume (Gbp) |
| <br>Cow 2  | <b>control</b><br>82 Gbp         | <b>3-NOP</b><br>118 Gbp          | <b>3-NOP + nitrate</b><br>82 Gbp | <b>nitrate</b><br>117 Gbp        | 1                  | 71                |
| <br>Cow 3  | <b>nitrate</b><br>113 Gbp        | <b>3-NOP + nitrate</b><br>99 Gbp | <b>3-NOP</b><br>99 Gbp           | <b>control</b><br>81 Gbp         | 2                  | 86                |
| <br>Cow 4 | <b>3-NOP</b><br>92 Gbp           | <b>nitrate</b><br>89 Gbp         | <b>control</b><br>112 Gbp        | <b>3-NOP + nitrate</b><br>91 Gbp | 3                  | 95                |
|                                                                                             |                                  |                                  |                                  |                                  | 4                  | 97                |
|                                                                                             |                                  |                                  |                                  |                                  | 5                  | 82                |
|                                                                                             |                                  |                                  |                                  |                                  | 6                  | 118               |
|                                                                                             |                                  |                                  |                                  |                                  | 7                  | 82                |
|                                                                                             |                                  |                                  |                                  |                                  | 8                  | 117               |
|                                                                                             |                                  |                                  |                                  |                                  | 9                  | 113               |
|                                                                                             |                                  |                                  |                                  |                                  | 10                 | 99                |
|                                                                                             |                                  |                                  |                                  |                                  | 11                 | 99                |
|                                                                                             |                                  |                                  |                                  |                                  | 12                 | 81                |
|                                                                                             |                                  |                                  |                                  |                                  | 13                 | 92                |
|                                                                                             |                                  |                                  |                                  |                                  | 14                 | 89                |
|                                                                                             |                                  |                                  |                                  |                                  | 15                 | 112               |
|                                                                                             |                                  |                                  |                                  |                                  | 16                 | 91                |

| DATA QUALITY                                                                                                             | 1     | 2     | 3     | 4     | 5     | 6     | 7     | 8     | 9     | 10    | 11    | 12    | 13    | 14    | 15    | 16    |
|--------------------------------------------------------------------------------------------------------------------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|
|  <b>N50</b><br>(Higher is better)     | 7634  | 7867  | 7823  | 9132  | 7751  | 8653  | 7445  | 8635  | 9086  | 8669  | 7421  | 8683  | 8176  | 9955  | 7676  | 7873  |
|  <b>Q score</b><br>(Higher is better) | 23.02 | 23.02 | 23.02 | 23.02 | 23.02 | 23.01 | 23.01 | 23.01 | 22.09 | 22.08 | 22.09 | 22.09 | 22.09 | 22.09 | 22.07 | 23.00 |

Treatments: control (gray) | nitrate (blue) | 3-NOP (yellow) | 3-NOP + nitrate (purple)

# Synergy with Microhub

- With support from Microhub bioinformatics team:
  - We are contributing to the methodology for taxonomy and functional quantification.
    - We are leading the part of generation database for genome-based taxonomy quantification.
  - We are leading the genome assembly part.
- All activities in MicroHub are valuable to improve our current work.

Thanks!

# Assigning Functional Annotation To Microbiome Reads

## Optimising Pipelines for Large-Scale Ruminant Genetics

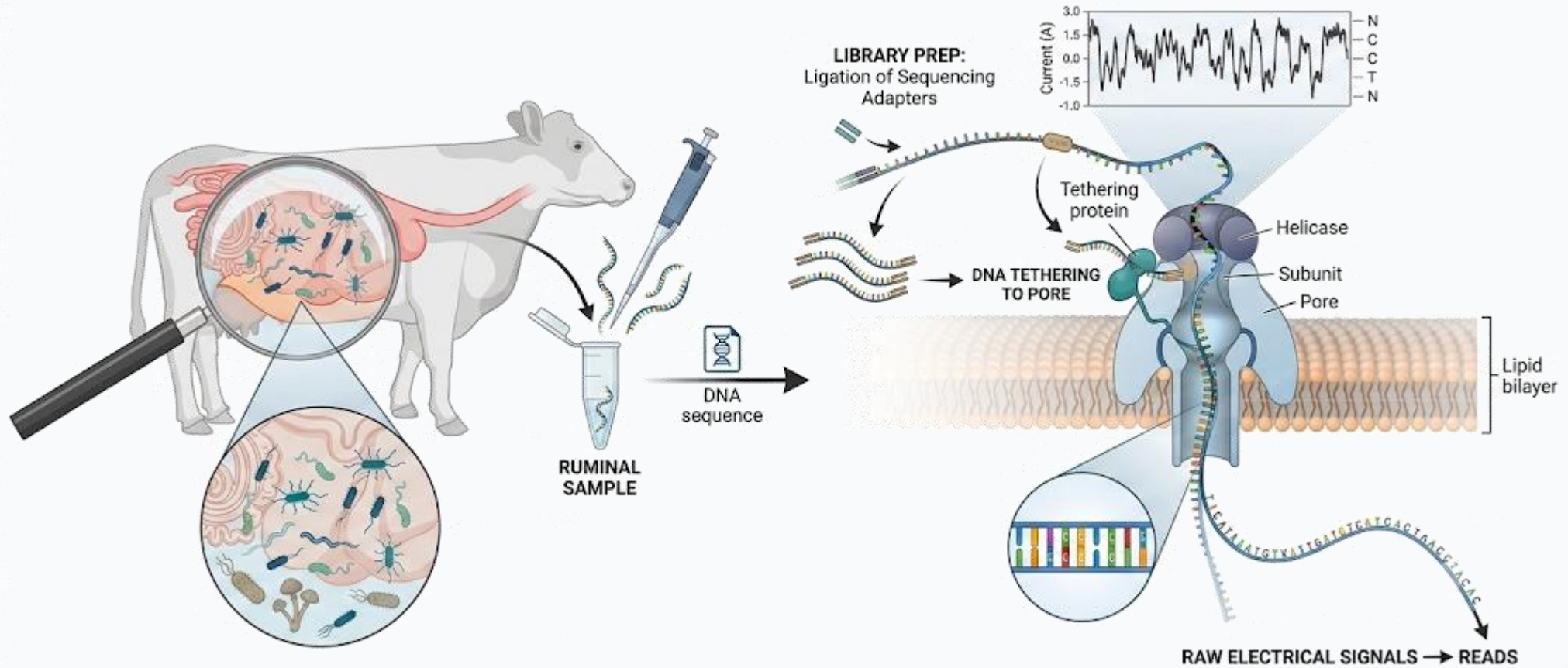


## Linking Microbiomes to Emissions

Our primary objective is mapping ruminal microbiomes to phenotypes of interest, specifically **methane emissions**.

By processing thousands of cattle ruminal samples through our streamlined functional pipeline, we isolate exact differential pathways (e.g. KEGG pathways) distinguishing high vs. low methane emitters.

# Sequencing the ruminal microbiome



# The Bioinformatic Fork



## Pathway A: Taxonomic Annotation

### WHO IS THERE?

Profiles the structural composition and species spectrum native to the rumen ecosystem

- ✓ **Reference Mapping:** Aligns raw reads directly against comprehensive genomic reference databases
- ✓ **Multi-Kingdom Detection:** Profiles Bacteria, Eukaryotes (protozoa/fungi), and Archaea simultaneously
- ✓ **Taxon Resolution:** Focuses on species-level classifications to identify exact names (e.g., *Methanobrevibacter ruminantium*)



## Pathway B: Functional Annotation

### WHAT ARE THEY DOING?

Deciphers the active metabolic potential and biochemical machinery within the microbiome

- ✓ **Enzymatic Pathways:** Maps genomic read signatures to chemical processes and pathways
- ✓ **KEGG Databases:** Aligns sequence reads to functional metabolic networks and structural biochemical pathways
- ✓ **COG Classifications:** Group proteins into broad, conserved evolutionary clusters to track core cellular mechanisms

# Why Functional Profiling Matters

## TAXONOMY ONLY

### "Who is there?"

Provides a simple descriptive catalog of species (e.g., *Escherichia coli* 15%)

- ❌ **Uncertain Capabilities:** Highly heritable names but unclear physical pathways or functional outputs
- ❌ **High Redundancy:** Species taxonomy varies wildly across herds, while chemical capability is identical
- ❌ **Weak Associations:** Suboptimal correlation with complex animal phenotypes (methane, feed intake)

## FUNCTIONAL PATHING

### "What are they doing?"

Provides mechanistic insight by directly measuring biological pathways and enzymes

- ✅ **Conserved Roles:** Focuses on active pathways (butyrate production, methane metabolism) despite shifts in individual taxons
- ✅ **Direct Selectability:** Measures biochemical targets directly affected by the host genome
- ✅ **Biological Model:** Uncovers antibiotic resistance, cellulose degradation, and core metabolism

# Our pipeline

SqueezeMeta: a fully automated metagenomics pipeline,  
from reads to bins

NEW: Now you can inspect the results of SqueezeMeta using our interactive viewer  
[Watermelon.](#)



`sqm_longreads.pl`

Taxonomy and  
functional annotation  
on the raw reads  
Diamond Blastx



`sqm_annot.pl`

Only functional  
annotation  
Prodigal

# The Computational Bottleneck



## Traditional Pipeline

The standard script **sqm\_longreads.pl** executes Diamond Blastx on the raw reads to acquire taxonomy & function simultaneously.

**50–60h** Per  
Sample

**40 Years** Total  
CPU Time



## The Streamlined Strategy

Splitting functional profiling and taxonomic profiling into separate dedicated lightweight steps via the custom script **sqm\_annot.pl**.

**2h** Func.  
Annot

**+2h** Taxonomy  
(Bracken)

# Faster Pipelines

## Over 90% Processing Time Saved

By utilizing decoupled pipelines, we avoid massive Diamond Blastx sequence space runs, which completely eliminates unnecessary processing overhead.

Scale calculation for 7,000 samples:

Standard Pipeline: 385,000 Core Hours

Decoupled Pipeline: 28,000 Core Hours

Traditional sqm\_longreads.pl

55 Hours / Sample

100% CPU Load



Decoupled Pipeline (sqm\_annot.pl + Kraken)

4 Hours / Sample

# Accelerated Pipeline Validation

## ACCURACY PROOF

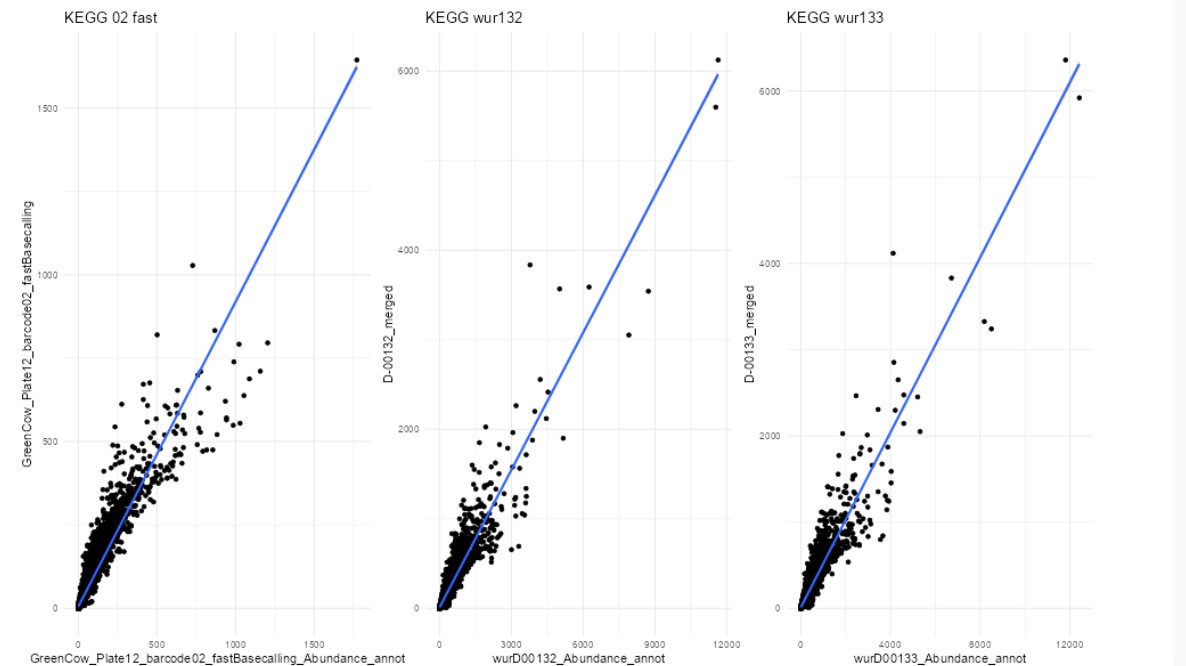
## Identical Biological Insight

Plotting relative abundance profiles derived from the full pipeline against the fast functional annotator results shows remarkable consistency

**Functional Correlation = 0.94**

Highly stable alignment was verified across COG and KEGG databases on multiple samples

*The fast pipeline delivers the exact same selection values with negligible variance*



# Mapping the Methane Phenotype

## DIFFERENTIAL ABUNDANCE

### Isolating Methane Biomarkers

To decipher pathways driving emissions, cows were ranked into quartile groups (Low, L-Mid, H-Mid, High) using Breath Infrared detectors

- ✓ **Linear Regression (Limma):** Identified 279 Differentially Abundant KEGGs (KEGGs-DA) with adjusted  $P < 0.05$
- ✓ **Emission Clusters:** 182 KEGGs linked to High Methane, and 97 KEGGs linked to Low Methane emitters
- ✓ **Functional PC1:** Aggregating KEGGs-DA explained 53.0% of functional metagenome variation

LIMMA LINEAR REGRESSION AND BENJAMINI-HOCHBERG CORRECTION



Journal of Dairy Science  
Volume 104, Issue 7, July 2021, Pages 8135-8151



Research

## A dimensional reduction approach to modulate the core ruminal microbiome associated with methane emissions via selective breeding

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Mónica Gutiérrez-Rivas <sup>1</sup>, Raquel Atxaerandio <sup>3</sup>, Idoia Goiri <sup>3</sup>, Aser  
García-Rodríguez <sup>3</sup>, José A. Jiménez-Montero <sup>4</sup>, Carmen González <sup>1</sup>,  
Javier Tamames <sup>5</sup>, Fernando Puente-Sánchez <sup>5</sup>, Luis Varona <sup>6</sup>,  
Magdalena Serrano <sup>1</sup>, Cristina Ovilo <sup>1</sup>, Oscar González-Recio <sup>1 7</sup>

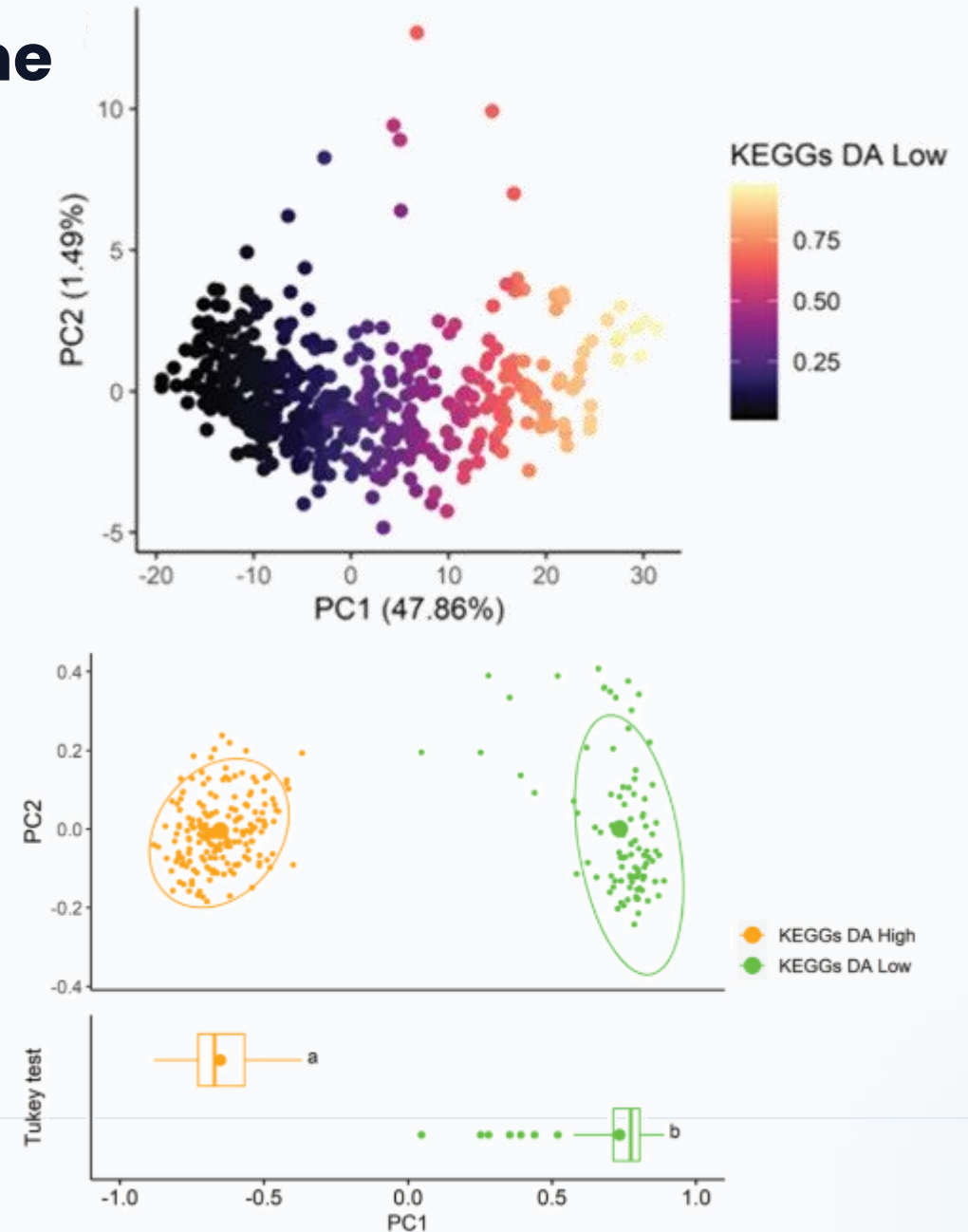
# Host Genomic Control of Microbiome

## HOLOGENE HERITABILITY

### The Core Rumen is Heritable

By running bivariate animal models with genomic relationship matrices, we proved the host genotype regulates the PC1 aggregated metagenome.

- ✓ **PC1 Heritabilities:** Consistent ~0.30 across all taxonomic levels.
- ✓ **Functional Heritability:** KEGGs-DA heritability reached
- ✓ **Direct Selectability:** Rumen properties are more heritable than physical methane measurements (~0.15)

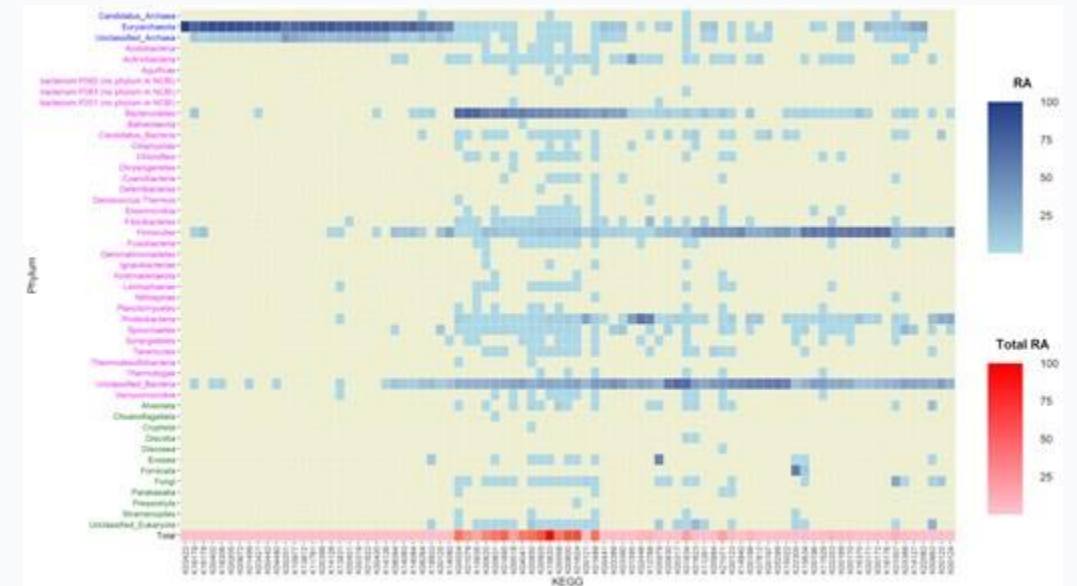


# Host Genomic Control of Microbiome

## We do not lose information

We can still know:

- ✓ What genes are more active in different taxa
- ✓ Keep track of what genera/species are associated with functions of interest
- ✓ Important to understand the micro-ecology



# Final ideas



## Dimension Reduction

Condensing thousands of differential KEGG pathway variables into structured Principal Components (PCs) for cleaner analysis.



## Heritability Mapping

Variations in these microbiome PCs are highly heritable and controlled by the host genome.



## Selective Breeding

Incorporating these host-microbiome genetic traits into national breeding indexes to select for low-emission cattle.



Thank you